

DAFTAR PUSTAKA

- Abramenko, N., Demidova, T. B., Krutyakov, Y. A., Zherebin, P. M., Krysanov, E. Y., Kustov, L. M., & Peijnenburg, W. (2019). The effect of capping agents on the toxicity of silver nanoparticles to *Danio rerio* embryos. *Nanotoxicology*, 1–13.
- Abu-Saieda, M. A. Tahab, E. Tarek, H. Nehal, & Elsayed. (2017). Polyvinyl alcohol/Sodium alginate integrated silver nanoparticles as probable solution for decontamination of microbes contaminated water. *International Journal of Biological Macromolecules*. 107. 10.1016/j.ijbiomac.2017.10.047.
- Ahmad, M. & Nofrizal. (2011). Pemijahan dan Penjinakkan Ikan Pantau (*Rasbora lateristriata*). *Jurnal Perikanan dan Kelautan*, 16 (1): 71–78.
- Al-Awady, M. J., Greenway, G. M., & Paunov, V. N. (2015). Nanotoxicity of polyelectrolyte-functionalized titania nanoparticles towards microalgae and yeast: role of the particle concentration, size and surface charge. *RSC Advances*, 5(46), 37044–37059. doi:10.1039/c5ra05577f
- Ašmonaitė, G., Boyer, S., Souza, K. B. de, Wassmur, B., & Sturve, J. (2016). Behavioural toxicity assessment of silver ions and nanoparticles on zebrafish using a locomotion profiling approach. *Aquatic Toxicology*, 173, 143–153. doi:10.1016/j.aquatox.2016.01.013
- Bai, C., & Tang, M. (2019). Toxicological study of metal and metal oxide nanoparticles in zebrafish. *Journal of Applied Toxicology*. doi:10.1002/jat.3910
- Bar-Ilan, O., Albrecht, R. M., Fako, V. E., & Furgeson, D. Y. (2009). Toxicity Assessments of Multisized Gold and Silver Nanoparticles in Zebrafish Embryos. *Small*, 5(16), 1897–1910. doi:10.1002/smll.200801716
- Benn, T., Cavanagh, B., Hristovski, K., Posner, J. D., & Westerhoff, P. (2010). The Release of Nanosilver from Consumer Products Used in the Home. *Journal of Environment Quality*, 39(6), 1875. doi:10.2134/jeq2009.0363
- Böhme, S., Stärk, H.-J., Reemtsma, T., & Kühnel, D. (2015). Effect propagation after silver nanoparticle exposure in zebrafish (*Danio rerio*) embryos: a correlation to internal concentration and distribution patterns. *Environmental Science: Nano*, 2(6), 603–614. doi:10.1039/c5en00118h
- Böhme, S., Baccaro, M., Schmidt, M., Potthoff, A., Stärk, H.-J., Reemtsma, T., & Kühnel, D. (2017). Metal uptake and distribution in the zebrafish (*Danio rerio*) embryo: differences between nanoparticles and metal ions. *Environmental Science: Nano*, 4(5), 1005–1015. doi:10.1039/c6en00440g
- Djumanto, E Setyobudi, AA Sentosa, R Budi, & N Nirwati. (2008). Reproductive Biology of the Yellow Rasbora (*Rasbora lateristriata*) Inhabitat of the Ngrancah River, Kulon Progo Regency. *Jurnal Perikanan (Journal of Fish Science)* 10 (2): 261-275.
- Djumanto & F. Setyawan. (2009). Food Habits of the Yellow Rasbora, *Rasbora lateristriata*, family: cyprinidae, Broodfish during Moving to Spawning Ground. *Jurnal Perikanan (Journal of Fish Science)* 11 (1): 107-114.

- Epilurahman, R, HA Asti, S Hadisusanto, DS Yudha, Trijoko, RS Ramadani, FXS Pranoto, IA Muhtianda. (2018). *Kekayaan Fauna Gianyar, Bali: Udang, Ikan, Amfibi, Reptil, Burung dan Mamalia*. Gadjah Mada University Press. Yogyakarta, p. 30.
- Hedberg, J., Oromieh, A. G., Kleja, D. B., & Wallinder, I. O. (2015). Sorption and dissolution of bare and coated silver nanoparticles in soil suspensions--Influence of soil and particle characteristics. *Journal of environmental science and health. Part A, Toxic/hazardous substances & environmental engineering*, 50(9), 891–900. <https://doi.org/10.1080/10934529.2015.1030271>
- Iravani, S., H. Korbekandi, S. V. Mirmohammadi, & B. Zolfaghari. (2014). Synthesis of silver nanoparticles: chemical, physical and biological methods. *Research in pharmaceutical sciences*. 9(6), 385–406.
- Kashiwada, S. (2006). Distribution of nanoparticles in the see-through medaka (*Oryzias latipes*). *Environ. Health Perspect.* 114, 1697–1702.
- Keller, A. A., McFerran, S., Lazareva, A., & Suh, S. (2013). Global life cycle releases of engineered nanomaterials. *Journal of Nanoparticle Research*, 15(6). doi:10.1007/s11051-013-1692-4
- Lee, K.J., Nallathamby, P.D., Browning, L.M., Osgood, C.J., Xu, X.H.N., (2007). In vivo imaging of transport and biocompatibility of single silver nanoparticles in early development of zebrafish embryos. *ACS Nano* 1, 133–143
- Lee, K. Y., & D. J. Mooney. (2012). Alginate: Properties and applications. *Progress in Polymer Science*. 37(1), 106–126.
- Liao, C., Li, Y., & Tjong, S. (2019). Bactericidal and Cytotoxic Properties of Silver Nanoparticles. *International Journal of Molecular Sciences*, 20(2), 449.
- Liu Y., S. Chen, L. Zhong, & G. Wu. (2009). Preparation of high-stable silver nanoparticle dispersion by using sodium alginate as a stabilizer under gamma radiation. *Radiat. Phys. Chem. Oxf. Engl* 1993. **78**: 251–255.
- Massarsky, A., Trudeau, V. L., & Moon, T. W. (2014). Predicting the environmental impact of nanosilver. *Environmental Toxicology and Pharmacology*, 38(3), 861–873. doi:10.1016/j.etap.2014.10.006
- Morones-Ramirez, J. R., J. A. Winkler, C. S. Spina, & J. J. Collins. (2013). Silver Enhances Antibiotic Activity Against Gram-Negative Bacteria. *Science Translational Medicine*. 5(190), 190ra81–190ra81. doi:10.1126/scitranslmed.3006276 .
- Mosselhy, D. A., He, W., Li, D., Meng, Y., & Feng, Q. (2016). Silver nanoparticles: in vivo toxicity in zebrafish embryos and a comparison to silver nitrate. *Journal of Nanoparticle Research*, 18(8). doi:10.1007/s11051-016-3514-y
- Nagasawa, N., H. Mitomo, F. Yoshii, and T. Kume (2000). Radiation-induced degradation of sodium alginate. *Polymer Degradation and Stability*. 69(3), 279–285. doi:10.1016/s0141-3910(00)00070-7.
- Naghavi, K., Saion, E., Rezaee, K., & Yunus, W. M. M. (2010). Influence of dose on particle size of colloidal silver nanoparticles synthesized by gamma radiation. *Radiation Physics and Chemistry*, 79(12), 1203–1208. doi:10.1016/j.radphyschem.2010.07.009

- Osborne, O. J., Johnston, B. D., Moger, J., Balousha, M., Lead, J. R., Kudoh, T., & Tyler, C. R. (2013). Effects of particle size and coating on nanoscale Ag and TiO₂ exposure in zebrafish (*Danio rerio*) embryos. *Nanotoxicology*, 7(8), 1315–1324. <https://doi.org/10.3109/17435390.2012.737484>
- Osborne, O. J., Lin, S., Chang, C. H., Ji, Z., Yu, X., Wang, X., ... Nel, A. E. (2015). Organ-Specific and Size-Dependent Ag Nanoparticle Toxicity in Gills and Intestines of Adult Zebrafish. *ACS Nano*, 9(10), 9573–9584. doi:10.1021/acsnano.5b04583
- Pareek, V., R. Gupta, & J. Panwar. (2018). Do physico-chemical properties of silver nanoparticles decide their interaction with biological media and bactericidal action? A review. *Materials Science and Engineering: C*, 90, 739–749. doi:10.1016/j.msec.2018.04.09.3
- Powers, C. M., Slotkin, T. A., Seidler, F. J., Badireddy, A. R., & Padilla, S. (2011). Silver nanoparticles alter zebrafish development and larval behavior: Distinct roles for particle size, coating and composition. *Neurotoxicology and Teratology*, 33(6), 708–714. doi:10.1016/j.ntt.2011.02.002
- Qin, Y., X., Jing, J., Liu, H., Wu, H., & Yang, W., (2010). Size control over spherical silver nanoparticle by ascorbic acid reduction. *Colloids Surfaces A Physicochem. Eng. Asp.* 372, 172-176
- Retnoaji B, Nanda F, Sartika D, Eunike N, Oktaviani DD, & Afriani D. (2016). The effect of volcanic dust on histological structure of wader pari (*Rasbora lateristriata* Bleeker, 1854) organs. *AIP Conference Proceedings*. 1744(1).
- Rigueur, D., & Lyons, K. M. (2014). *Whole-Mount Skeletal Staining. Skeletal Development and Repair*, 113–121. doi:10.1007/978-1-62703-989-5_9
- Santoshkumar S, V., & S, R. (2019). Toxicology evaluation and Antidermatophytic activity of silver nanoparticles synthesized using leaf extract of *Passiflora caerulea*. *South African Journal of Chemical Engineering*. doi:10.1016/j.sajce.2019.04.001
- Schluesener, J. K., & Schluesener, H. J. (2013). Nanosilver: application and novel aspects of toxicology. *Archives of Toxicology*, 87(4), 569–576. doi:10.1007/s00204-012-1007-z
- Sheikh, N., A. Akhavan, & M. Z. Kassae. (2009). Synthesis of antibacterial silver nanoparticles by γ -irradiation. *Physica E: Low-Dimensional Systems and Nanostructures*. 42(2), 132–135. doi:10.1016/j.physe.2009.09.013
- Syafiuddin, A., Salmiati, Salim, M. R., Beng Hong Kueh, A., Hadibarata, T., & Nur, H. (2017). A Review of Silver Nanoparticles: Research Trends, Global Consumption, Synthesis, Properties, and Future Challenges. *Journal of the Chinese Chemical Society*, 64(7), 732–756. doi:10.1002/jccs.201700067
- Vance, M. E., Kuiken, T., Vejerano, E. P., McGinnis, S. P., Hochella, M. F., Rejeski, D., & Hull, M. S. (2015). Nanotechnology in the real world: Redeveloping the nanomaterial consumer products inventory. *Beilstein Journal of Nanotechnology*, 6, 1769–1780. doi:10.3762/bjnano.6.181
- Velgosova, O., Čížmárová, E., Málek, J., & Kavuličova, J. (2017). Effect of storage conditions on long-term stability of Ag nanoparticles formed via green

- synthesis. *International Journal of Minerals, Metallurgy, and Materials*, 24(10), 1177–1182. doi:10.1007/s12613-017-1508-0
- Weigele, J., & Franz-Odenaal, T. A. (2016). Functional bone histology of zebrafish reveals two types of endochondral ossification, different types of osteoblast clusters and a new bone type. *Journal of Anatomy*, 229(1), 92–103. doi:10.1111/joa.12480
- Wu, Y., Zhou, Q., Li, H., Liu, W., Wang, T., & Jiang, G. (2010). Effects of silver nanoparticles on the development and histopathology biomarkers of Japanese medaka (*Oryzias latipes*) using the partial-life test. *Aquatic Toxicology*, 100(2), 160–167. doi:10.1016/j.aquatox.2009.11.014
- Wu, Y., & Zhou, Q. (2012). Dose- and time-related changes in aerobic metabolism, chorionic disruption, and oxidative stress in embryonic medaka (*Oryzias latipes*): underlying mechanisms for silver nanoparticle developmental toxicity. *Aquatic toxicology (Amsterdam, Netherlands)*, 124-125, 238–246. <https://doi.org/10.1016/j.aquatox.2012.08.009>