DAFTAR PUSTAKA

Adagba, O., Walet Pierre, G., Kouamé François, G., Kouassi Badama Philomène, S., Louis Rois Nondenot, A., Romain, A., & Ismael, K. (2015). Inventory of the mealybug species associated to the cocoa tree (Theobroma cacao L.) in four producing areas infected with the swollen shoot disease in Côte d'Ivoire. *~ 312 ~ Journal of Entomology and Zoology Studies*, *3*(4), 312–316.

Aimone, C. D., Hoyer, J. S., Dye, A. E., Deppong, D. O., Duffy, S., Carbone, I., & Hanley-Bowdoin, L. (2022). An experimental strategy for preparing circular ssDNA virus genomes for next-generation sequencing. *Journal of Virological Methods*, *300*. https://doi.org/10.1016/j.jviromet.2021.114405

Alif, T., Hartono, S., & Sulandari, S. (2018). Karakterisasi Virus Penyebab Penyakit Belang pada Tanaman Lada (Piper nigrum L.). *Jurnal Perlindungan Tanaman Indonesia*, *22*(1), 115. https://doi.org/10.22146/jpti.30354

Ameyaw, G. A., Chingandu, N., Domfeh, O., Dzahini-Obiatey, H. K., Gutierrez, O. A., & Brown, J. K. (2017). Variable detection of Cacao swollen shoots disease-associated badnaviruses by PCR amplification. *International Symposium of Cocoa Research (ISCR)*, 13–17.

Ameyaw, G. A., Domfeh, O., Armooh, B., Boakye, A. Y., & Arjarquah, A. (2022). Inconsistent PCR detection of Cacao swollen shoot virus (CSSV) is linked to the occurrence of different variants across the cocoa regions of Ghana. *Journal of Virological Methods*, *300*. https://doi.org/10.1016/j.jviromet.2021.114400

Ariningsih, E., Pakpahan, A., Sinuraya, J. F., Purba, H. J., Suharyono, S., & Septanti, K. S. (2020). Increasing added value of superior agricultural products. In *Agro-Socioeconomic Newsletter* (Vol. 13, Issue 3, pp. 1–3).

Asrul, L., Yulianti, A., Musa, Y., & Trisnaputri, A. C. (2021). The Development Strategies of Cocoa Cultivation (Theobroma cacao L.) in Soppeng District. *IOP Conference Series: Earth and Environmental Science*, *921*(1). https://doi.org/10.1088/1755-1315/921/1/012050

Badan Pusat Statistik. (2022). *Statistik Kakao Indonesia 2021*. Jakarta

Bhat, A. I., Hohn, T., & Selvarajan, R. (2016). Badnaviruses: The current global scenario. In *Viruses* (Vol. 8, Issue 6). MDPI AG. https://doi.org/10.3390/v8060177

Bhattacharjee, R., & Akoroda, M. (2018). *Taxonomy and classification of cacao* (pp. 3–18). https://doi.org/10.19103/as.2017.0021.01

Bryceson, S. R., Morgan, J. W., McMahon, P. J., & Keane, P. J. (2023). A sudden and widespread change in symptoms and incidence of vascular streak dieback of cocoa (Theobroma cacao) linked to environmental change in Sulawesi, Indonesia. *Agriculture, Ecosystems and Environment*, *350*. https://doi.org/10.1016/j.agee.2023.108466

CABI. (2019a). *Penyakit kakao : Panduan Foto dan Hama Penyakit dari CABI*. CABI-Plantwise. https://cabidigitallibrary.org

CABI. (2019b). *Theobroma cacao*. https://www.cabidigitallibrary.org/. Diakses pada 02 Januari 2021

Campbell, C. A. M. (1983). The assessment of mealybugs (Pseudococcidae) and other Homoptera on mature cocoa trees in Ghana. *Bulletin of Entomological Research*, *73*(1), 137–151. https://doi.org/10.1017/S0007485300013870

Cenis, J. L., Perez, P., & Fereres, A. (1993). Identification of Aphid (Homoptera: Aphididae) Species and Clones by Random Amplified Polymorphic DNA. *Annals of the Entomological Society of America*, *86*(5), 545–550. https://doi.org/10.1093/aesa/86.5.545

Charles, J. G. (2011). Using parasitoids to infer a native range for the obscure mealybug, Pseudococcus viburni, in South America. *BioControl*, *56*, 155–161. https://doi.org/10.1007/s10526-010-9322-x

Chingandu, N., Dongo, L., Gutierrez, O. A., & Brown, J. K. (2019). The previously unidentified, divergent badnavirus species cacao red vein-banding virus is associated with cacao swollen shoot disease in Nigeria. *Plant Disease*, *103*(6), 1302–1308. https://doi.org/10.1094/PDIS-09-18-1561-RE

Chingandu, N., Kouakou, K., Aka, R., Ameyaw, G., Gutierrez, O. A., Herrmann, H. W., & Brown, J. K. (2017). The proposed new species, cacao red vein virus, and three previously recognized badnavirus species are associated with cacao swollen shoot disease. *Virology Journal*, *14*(1). https://doi.org/10.1186/s12985-017-0866-6

Chingandu, N., Zia-ur-rehman, M., Sreenivasan, T. N., Surujdeo-Maharaj, S., Umaharan, P., Gutierrez, O. A., & Brown, J. K. (2017). Molecular characterization of previously elusive badnaviruses associated with symptomatic cacao in the New World. *Archives of Virology*, *162*(5), 1363–1371. https://doi.org/10.1007/s00705-017-3235-2

DAFF Australia. (2019). *Final group pest risk analysis for mealybugs and the viruses they transmit on fresh fruit, vegetable, cut-flower and foliage imports*. Australia.

de Tomás, C., & Vicient, C. M. (2022). Genome-wide identification of Reverse Transcriptase domains of recently inserted endogenous plant pararetrovirus (Caulimoviridae). *Frontiers in Plant Science*, *13*. https://doi.org/10.3389/fpls.2022.1011565

Dewi Hs, E. S., Yudono, P., Putra, E. T. S., & Purwanto, B. H. (2020). Physiological and biochemical activities of cherelle wilt on three cocoa clones (Theobroma cacao) under two levels of soil fertilities. *Biodiversitas*, *21*(1), 187–194. https://doi.org/10.13057/biodiv/d210124

Ding, S. W., Mackenzie, A., Torronen, M., & Gibbs, A. (1990). Nucleotide sequence of the virion protein gene of cacao yellow mosaic tymovirus. *Nucleic Acids Research*, *18*(19), 5886. https://doi.org/10.1093/nar/18.19.5886

Dzahini-Obiatey, H., Domfeh, O., & Amouah, F. M. (2010). Over seventy years of a viral disease of cocoa in Ghana: From researchers' perspective. In *African Journal of*

Agricultural Research (Vol. 5, Issue 7). https://www.researchgate.net/publication/267683769

Effendy, Fardhal Pratama, M., Rauf, R. A., Antara, M., Basir-Cyio, M., Mahfudz, & Muhardi. (2019). Factors influencing the efficiency of cocoa farms: A study to increase income in rural Indonesia. *PLoS ONE*, *14*(4). https://doi.org/10.1371/JOURNAL.PONE.0214569

End, M. J., Daymond, A. J., & Hadley, P. (2021). *Technical Guidelines for the Safe Movement of Cacao Germplasm*. Global Cacao Genetic Resources Network (CacaoNet), Bioversity International (CacaoNet). www.cacaonet.org

FAOSTAT. (2020). *FAOSTAT : Cocoa Bean Production*. https://data.un.org/Data.aspx?d=FAO&f=itemCode%3A661. Diakses pada 2 Januari 2021

Folmer, O., Black, M., Hoeh, W., Lutz, R., & Vrijenhoek, R. (1994). DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. In *Molecular Marine Biology and Biotechnology* (Vol. 3, Issue 5).

Furuya, N., Suastika, G., & Natsuaki, K. T. (2012). First Report and Molecular Characterization of Exogenous Banana Steak Mysore Virus from Banana in Indonesia. *Asian Journal of Plant Pathology*, *6*(2), 41–47.

Garcia-Morales, M., Denno, D. B., Miller, D. R., Miller, G. L., Ben-Dov, Y., & Hardy, N. B. (2016, February 9). *ScaleNet: a literature-based model of scale insect biology and systematics*. Database. https://doi.org/10.1093/database/bav118

Gayral, P., Noa-Carrazana, J.-C., Lescot, M., Lheureux, F., Lockhart, B. E. L., Matsumoto, T., Piffanelli, P., & Iskra-Caruana, M.-L. (2008). A Single Banana Streak Virus Integration Event in the Banana Genome as the Origin of Infectious Endogenous Pararetrovirus . *Journal of Virology*, *82*(13), 6697–6710. https://doi.org/10.1128/jvi.00212-08

Geering, A. D. W., Scharaschkin, T., & Teycheney, P. Y. (2010). The classification and nomenclature of endogenous viruses of the family Caulimoviridae. *Archives of Virology*, *155*(1), 123–131. https://doi.org/10.1007/s00705-009-0488-4

Guenoune-Gelbart, D., Sufrin-Ringwald, T., Capobianco, H., Gaba, V., Polston, J. E., & Lapidot, M. (2010). Inoculation of plants with begomoviruses by particle bombardment without cloning: Using rolling circle amplification of total DNA from infected plants and whiteflies. *Journal of Virological Methods*, *168*(1–2), 87–93. https://doi.org/10.1016/j.jviromet.2010.04.022

Guest, D., & Keane, P. (2007). Vascular-streak dieback: A new encounter disease of cacao in Papua New Guinea and Southeast Asia caused by the obligate basidiomycete Oncobasidium theobromae. *Phytopathology*, *97*(12), 1654–1657. https://doi.org/10.1094/PHYTO-97-12-1654

Gyamera, E. A., Domfeh, O., & Ameyaw, G. A. (2023). Cacao Swollen Shoot Viruses in Ghana. *Plant Disease*, *107*(5), 1261–1278. https://doi.org/10.1094/PDIS-10-22-2412-FE

Harper, G., Hull, R., Lockhart, B., & Olszewski, N. (2002). Viral sequences integrated into plant genomes. In *Annual Review of Phytopathology* (Vol. 40, pp. 119–136). https://doi.org/10.1146/annurev.phyto.40.120301.105642

Hohn, T., & Rothnie, H. (2013). Plant pararetroviruses: Replication and expression. In *Current Opinion in Virology* (Vol. 3, Issue 6, pp. 621–628). Elsevier B.V. https://doi.org/10.1016/j.coviro.2013.08.013

Hull, R. (2001). Classifying reverse transcribing elements: a proposal and a challenge to the ICTV. In *Arch Virol* (Vol. 146). Virology Division News.

Jahn, G. C., Beardsley, J. W., & González-Hernández, H. (2003). A Review of the Association of Ants with Mealybug Wilt Disease of Pineapple. *Proc. Hawaiian Entomol. Soc.*, *7777*(2), 9–28.

Jumanto, & Sumardiyono, Y. B. (1976). Beberapa pengamatan gejala mosaik pada daun coklat (Theobroma cacao L.) di kebun Beji dan Jatirunggo. *Kongres Nasional PFI Ke IV, Bandung, 20-21 Desember 1976*.

Kandito, A., Hartono, S., Trisyono, Y. A., & Somowiyarjo, S. (2022). First report of Cacao mild mosaic virus associated with cacao mosaic disease in Indonesia. In *New Disease Reports* (Vol. 45, Issue 2). John Wiley and Sons Inc. https://doi.org/10.1002/ndr2.12071

Kementerian Pertanian. (2020). *Statistik Pertanian 2019*. Jakarta

Kenten, R. H., & Woods, R. O. (1976). A virus of the cocoa swollen shoot group infecting cocoa in north sumatra. *PANS*, *22*(4), 488–490. https://doi.org/10.1080/09670877609414338

Kouakou, K., Kébé, B. I., Kouassi, N., Aké, S., Cilas, C., & Muller, E. (2012). Geographical distribution of Cacao swollen shoot virus molecular variability in Côte d'Ivoire. *Plant Disease*, *96*(10), 1445–1450. https://doi.org/10.1094/PDIS-09-11-0749-RE

Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, *33*(7), 1870–1874. https://doi.org/10.1093/molbev/msw054

Kuriyama, K., Tabara, M., Moriyama, H., Kanazawa, A., Koiwa, H., Takahashi, H., & Fukuhara, T. (2020). Disturbance of floral colour pattern by activation of an endogenous pararetrovirus, petunia vein clearing virus, in aged petunia plants. *Plant Journal*, *103*(2), 497–511. https://doi.org/10.1111/tpj.14728

Legg, J. T. (1977). *Cocoa Swollen Shoot Disease-Know Your Enemy*. Cocoa Research Institute of Ghana. Taifo, Ghana

Marelli, J. P., Guest, D. I., Bailey, B. A., Evans, H. C., Brown, J. K., Junaid, M., Barreto, R. W., Lisboa, D. O., & Puig, A. S. (2019). Chocolate under threat from old and new cacao diseases. In *Phytopathology* (Vol. 109, Issue 8, pp. 1331–1343). American Phytopathological Society. https://doi.org/10.1094/PHYTO-12-18-0477-RVW

Martel, N., Gomes, S. A., Chemin, I., Trépo, C., & Kay, A. (2013). Improved rolling circle amplification (RCA) of hepatitis B virus (HBV) relaxed-circular serum DNA (RC-DNA).

Journal of Virological Methods, *193*(2), 653–659. https://doi.org/10.1016/j.jviromet.2013.07.045

Menteri Pertanian Republik Indonesia. (2020). *Peraturan Menteri Pertanian no 25 tentang Organisme Pengganggu Tumbuhan Karantina.*

Mette, M. F., Kanno, T., Aufsatz, W., Jakowitsch, J., van der Winden, J., Matzke, M. A., & Matzke, A. J. M. (2002). Endogenous viral sequences and their potential contribution to heritable virus resistance in plants. *The EMBO Journal*, *21*(3), 461–469.

Motamayor, J. C., Lachenaud, P., da Silva e Mota, J. W., Loor, R., Kuhn, D. N., Brown, J. S., & Schnell, R. J. (2008). Geographic and genetic population differentiation of the Amazonian chocolate tree (Theobroma cacao L). *PLoS ONE*, *3*(10). https://doi.org/10.1371/journal.pone.0003311

Muhardi, Rahim, A., Effendy, Antara, M., Rauf, R. A., Lamusa, A., Christoporus, Hadayani, Safitri, D., & Mulyo, J. H. (2020). Sustainability of cocoa production in Indonesia. *Australian Journal of Crop Science*, *14*(6), 997–1003. https://doi.org/10.21475/ajcs.20.14.06.p2510

Muhire, B. M., Varsani, A., & Martin, D. P. (2014). SDT: A virus classification tool based on pairwise sequence alignment and identity calculation. *PLoS ONE*, *9*(9). https://doi.org/10.1371/journal.pone.0108277

Mujiono, T., & Samuji. (2005). Dampak Pengendalian Hama Terpadu Kakao terhadap Serangan Penggerek Buah Kakao (<i>Conopomorpha cramella<i> (Sn.))(Lepidoptera : Gracillariidae) dan <i>Helopeltis<i> sp. (Hemiptera : Miridae). *Jurnal Entomologi Indonesia*, *2*(1), 34–42.

Muller, E. (2016). Cacao swollen shoot virus (CSSV): History, biology, and genome. In *Cacao Diseases: A History of Old Enemies and New Encounters* (pp. 337–358). Springer International Publishing. https://doi.org/10.1007/978-3-319-24789-2_10

Muller, E., Ravel, S., Agret, C., Abrokwah, F., Dzahini-Obiatey, H., Galyuon, I., Kouakou, K., Jeyaseelan, E. C., Allainguillaume, J., & Wetten, A. (2018a). Next generation sequencing elucidates cacao badnavirus diversity and reveals the existence of more than ten viral species. *Virus Research*, *244*, 235–251. https://doi.org/10.1016/j.virusres.2017.11.019

Muller, E., Ravel, S., Agret, C., Abrokwah, F., Dzahini-Obiatey, H., Galyuon, I., Kouakou, K., Jeyaseelan, E. C., Allainguillaume, J., & Wetten, A. (2018b). Next generation sequencing elucidates cacao badnavirus diversity and reveals the existence of more than ten viral species. *Virus Research*, *244*, 235–251. https://doi.org/10.1016/j.virusres.2017.11.019

Muller, E., Ullah, I., Dunwell, J. M., Daymond, A. J., Richardson, M., Allainguillaume, J., & Wetten, A. (2021). Identification and distribution of novel badnaviral sequences integrated in the genome of cacao (Theobroma cacao). *Scientific Reports*, *11*(1). https://doi.org/10.1038/s41598-021-87690-1

Neilson, J., Dwiartama, A., Fold, N., & Permadi, D. (2020). Resource-based industrial policy in an era of global production networks: Strategic coupling in the Indonesian

cocoa sector. *World Development*, *135*. https://doi.org/10.1016/j.worlddev.2020.105045

Obok, E., Wetten, A., & Allainguillaume, J. (2018). Molecular Evidence of Cacao Swollen Shoot Virus Acquisition and Retention by Planococcus Citri (Risso) and Pseudococcus Longispinus (Targioni-Tozzetti) and Pseudococcus Viburni (Signoret) Mealybugs (Hemiptera: Pseudococcidae). *International Journal of Scientific and Research Publications (IJSRP)*, *8*(3). https://doi.org/10.29322/ijsrp.8.3.2018.p7507

Opoku, I. Y., Dakwa, J. T., & Ollennu, L. (2005). The development of Phytophthora canker on different cocoa genotypes infected and uninfected with cocoa swollen shoot virus. *Tropical Science* , *45*(1), 50–53.

Oro, F., Mississo, E., Okassa, M., Guilhaumon, C., Fenouillet, C., Cilas, C., & Muller, E. (2012). Geographical differentiation of the molecular diversity of cacao swollen shoot virus in Togo. *Archives of Virology*, *157*(3), 509–514. https://doi.org/10.1007/s00705-011-1158-x

Padi, F. K., Domfeh, O., Takrama, J., & Opoku, S. Y. (2013). An evaluation of gains in breeding for resistance to the cocoa swollen shoot virus disease in Ghana. *Crop Protection*, *51*, 24–31. https://doi.org/10.1016/j.cropro.2013.04.007

Pancaningtyas, S., & Wahyu Susilo, A. (2022). Analysis of Cocoa Clonal Seedlings Purity Through Deoxyribonucleic Acid (DNA) Barcoding and Random Amplification of Polymorphic DNA (RAPD) Fingerprinting. *Pelita Perkebunan (a Coffee and Cocoa Research Journal)*, *38*(1), 20–28. https://doi.org/10.22302/iccri.jur.pelitaperkebunan.v38i1.490

Park, D.-S., Leem, Y. J., Hahn, K.-W., Suh, S.-J., Hong, K.-J., & Oh, H.-W. (2010). Molecular Identification of Mealybugs (Hemiptera: Pseudococcidae) Found on Korean Pears. *J. Econ. Entomol*, *103*(1), 25–33. https://doi.org/10.1603/EC09144

Parnata, Y. (1976). Beberapa Catatan Mengenai Penyakit Virus Tanaman Coklat di Sumatra Utara. *Buletin BPP Medan*, *7*, 5–13.

Passricha, N., Saifi, S., Khatodia, S., & Tuteja, N. (2016). Assessing zygosity in progeny of transgenic plants: current methods and perspectives. *Journal of Biological Methods*, *3*(3), e46. https://doi.org/10.14440/jbm.2016.114

Pooggin, M. M. (2013). How can plant DNA viruses evade siRNA-directed DNA methylation and silencing? In *International Journal of Molecular Sciences* (Vol. 14, Issue 8, pp. 15233–15259). https://doi.org/10.3390/ijms140815233

Prawoto, & Martini. (2014). *Pedoman Budi Daya Kakao pada Kebun Campur*. Pusat Penelitian Kopi dan Kakao.

Probowati, W., Somowiyarjo, S., & Hartono, S. (2019). Molecular characterization of mosaic virus from the cocoa trees showing mosaic symptoms in yogyakarta, Indonesia. *Biodiversitas*, *20*(12), 3698–3704. https://doi.org/10.13057/biodiv/d201232

Puig, A. S. (2021a). Detection of cacao mild mosaic virus (CaMMV) using nested PCR and evidence of uneven distribution in leaf tissue. *Agronomy*, *11*(9). https://doi.org/10.3390/agronomy11091842

Puig, A. S. (2021b). Seed Transmission of a Cacao Virus From the Americas and the Implication on Crop Cultivation and Movement of Germplasm. *Plant Health Conference*. https://doi.org/10.13140/RG.2.2.19587.43045

Puig, A. S., Ramos-Sobrinho, R., Keith, C., Kitchen, N., Gutierrez, O., Goenaga, R., & Brown, J. K. (2020). First report of cacao mild mosaic virus associated with symptomatic commercial cacao (theobroma cacao) trees in puerto rico. In *Plant Disease* (Vol. 104, Issue 11, p. 3089). American Phytopathological Society. https://doi.org/10.1094/PDIS-04-20-0745-PDN

Puig, A. S., Wurzel, S., Suarez, S., Marelli, J. P., & Niogret, J. (2021). Mealybug (Hemiptera: Pseudococcidae) species associated with cacao mild mosaic virus and evidence of virus acquisition. *Insects*, *12*(11). https://doi.org/10.3390/insects12110994

Pusat Penelitian Kopi dan Kakao. (2015). *Pedoman Teknis Budidaya Tanaman Kakao*. Jember.

Quainoo, A. K., Wetten, A. C., & Allainguillaume, J. (2008a). The effectiveness of somatic embryogenesis in eliminating the cocoa swollen shoot virus from infected cocoa trees. *Journal of Virological Methods*, *149*(1), 91–96. https://doi.org/10.1016/j.jviromet.2008.01.007

Quainoo, A. K., Wetten, A. C., & Allainguillaume, J. (2008b). Transmission of cocoa swollen shoot virus by seeds. *Journal of Virological Methods*, *150*(1–2), 45–49. https://doi.org/10.1016/j.jviromet.2008.03.009

Ramos-Sobrinho, R., Chingandu, N., Gutierrez, O. A., Marelli, J. P., & Brown, J. K. (2020). A complex of badnavirus species infecting cacao reveals mixed infections, extensive genomic variability, and interspecific recombination. *Viruses*, *12*(4). https://doi.org/10.3390/v12040443

Ramos-Sobrinho, R., Ferro, M. M. M., Nagata, T., Puig, A. S., Keith, C. von, Britto, D. S., Gutierrez, O. A., Marelli, J.-P., & Brown, J. K. (2021). Complete genome sequences of three newly discovered cacao mild mosaic virus isolates from Theobroma cacao L. in Brazil and Puerto Rico and evidence for recombination. *Archives of Virology*, *166*(7), 2027–2031. https://doi.org/10.1007/s00705-021-05063-5

Richert-Pöggeler, K. R., & Minarovits, J. (2014). Diversity of latent plant–virus interactions and their impact on the virosphere. *Plant Virus-Host Interaction: Molecular Approaches and Viral Evolution*, 263–275. https://doi.org/10.1016/B978-0-12-411584-2.00014-7

Richert-Poggeler, K. R., Noreen, F., Schwarzacher, T., Harper, G., & Hohn, T. (2003). Induction of infectious petunia vein clearing (pararetro) virus from endogenous provirus in petunia. *The EMBO Journal*, *22*(18), 4836–4845.

Sartiami, D., Saptayanti, N., Syahputra, E., & Puji Mardiasih, W. (2020). Mealybugs (Hemiptera: Pseudococcidae) Associated with Dragon Fruit in Indonesia. In *Advances in Biological Sciences Research* (Vol. 8).

Sartiami, D., & Watson, G. W. (2017). *A taxonomic update of takahashi's historic collection of mealybugs (Hemiptera: Pseudococcidae) from Malaysia And Singapore*. https://www.researchgate.net/publication/323186203

Saunders, J. A., Mischke, S., Leamy, E. A., & Hemeida, A. A. (2004). Selection of international molecular standards for DNA fingerprinting of Theobroma cacao. *Theoretical and Applied Genetics*, *110*(1), 41–47. https://doi.org/10.1007/s00122-004-1762-1

Semangun, H. (1961). Gedjala-gedjala mosaik kakao. *Universitas Gadjah Mada*. Yogyakarta

Sether, D. M., Ullman, D. E., & Hu, J. S. (1998). Transmission of Pineapple Mealybug Wilt-Associated Virus by Two Species of Mealybug *(Dysmicoccus spp.)*. *Virology 86(11) 1224-1230*

Silva, V. C. P. da, Nondillo, A., Galzer, E. C. W., Garcia, M. S., & Botton, M. (2017). Effect of Host Plants on the Development, Survivorship, and Reproduction of Pseudococcus viburni (Hemiptera: *Pseudococcidae*). *Florida Entomologist*, *100*(4), 718–724. https://doi.org/10.1653/024.100.0418

Somowiyarjo, S., Sulandari, S., Hartono, S., Paradisa, Y. B., & Aji, T. M. (2014). Etiology of Shoot Twig Malformation on Cocoa at Kulon Progo, Special Province of Yogyakarta And Segayung, Central Java. *Jurnal Perlindungan Tanaman Indonesia*, *18*(2), 95–102.

Somowiyarjo, S., Sulandari, S., Hartono, S., Paradisa, Y. B., Tri, &, & Aji, M. (2014). *Etiology of Shoot Twig Malformation on Cocoa at Kulon Progo, Special Province of Yogyakarta And Segayung, Central Java*.

Staginnus, C., Gregor, W., Mette, M. F., Chee, H. T., Borroto-Fernández, E. G., Da Câmara Machado, M. L., Matzke, M., & Schwarzacher, T. (2007). Endogenous pararetroviral sequences in tomato (Solanum lycopersicum) and related species 24. *BMC Plant Biology*, *7*. https://doi.org/10.1186/1471-2229-7-24

Staginnus, C., & Richert-Pöggeler, K. R. (2006). Endogenous pararetroviruses: two-faced travelers in the plant genome. In *Trends in Plant Science* (Vol. 11, Issue 10, pp. 485–491). https://doi.org/10.1016/j.tplants.2006.08.008

Sukal, A. C., Kidanemariam, D. B., Dale, J. L., Harding, R. M., & James, A. P. (2019). Assessment and optimization of rolling circle amplification protocols for the detection and characterization of badnaviruses. *Virology*, *529*, 73–80. https://doi.org/10.1016/j.virol.2019.01.013

Syauqi, A., Dadang, D., Harahap, I. S., & Indarwatmi, M. (2021). Gamma irradiation against mealybug Dysmicoccus lepelleyi (Betrem) (Hemiptera: Pseudococcidae) on mangosteen fruit (Garcinia mangostana L.) as a quarantine treatment. *Radiation Physics and Chemistry*, *179*. https://doi.org/10.1016/j.radphyschem.2020.108954

Takahashi, H., Fukuhara, T., Kitazawa, H., & Kormelink, R. (2019). Virus Latency and the Impact on Plants. In *Frontiers in Microbiology* (Vol. 10). Frontiers Media S.A. https://doi.org/10.3389/fmicb.2019.02764

Teycheney, P. Y., Geering, A. D. W., Dasgupta, I., Hull, R., Kreuze, J. F., Lockhart, B., Muller, E., Olszewski, N., Pappu, H., Pooggin, M. M., Richert-Pöggeler, K. R., Schoelz, J. E., Seal, S., Stavolone, L., & Umber, M. (2020). ICTV Virus taxonomy profile: Caulimoviridae. In *Journal of General Virology* (Vol. 101, Issue 10, pp. 1025–1026). Microbiology Society. https://doi.org/10.1099/jgv.0.001497

Thorburn, C. (2015). The rise and demise of integrated pest management in rice in Indonesia. *Insects*, *6*(2), 381–408. https://doi.org/10.3390/insects6020381

Tomonaga, K., Suzuki, N., & Berkhout, B. (2019). “Integration of viral sequences into eukaryotic host genomes: legacy of ancient infections.” In *Virus Research* (Vol. 262, p. 1). Elsevier B.V. https://doi.org/10.1016/j.virusres.2018.12.012

Triharso, Sumardiyono, Y. B., Jumanto, & Soemanto. (1981). Studies on mosaic disease on cocoa in Indonesia. *International Conference on Tropical Crop Protection*.

Tyasningsiwi, R. W., Witjaksono, W., & Indarti, S. (2019). Analysis of Volatile Compound at Different Age of Corn Crops Used as Bemisia tabaci Repellent. *Jurnal Perlindungan Tanaman Indonesia*, *23*(1), 142. https://doi.org/10.22146/jpti.35954

Ullah, I., Daymond, A. J., Hadley, P., End, M. J., Umaharan, P., & Dunwell, J. M. (2021). Identification of cacao mild mosaic virus (Cammv) and cacao yellow vein-banding virus (cyvbv) in cocoa (theobroma cacao) germplasm. *Viruses*, *13*(11). https://doi.org/10.3390/v13112152

Ullah, I., & Dunwell, J. M. (2023). Bioinformatic, genetic and molecular analysis of several badnavirus sequences integrated in the genomes of diverse cocoa (Theobroma cacao L.) germplasm. *Saudi Journal of Biological Sciences*, 103648. https://doi.org/10.1016/j.sjbs.2023.103648

Ullah, I., Kamran, M., & Dunwell, J. M. (2023). Identification of a Novel Polerovirus in Cocoa (Theobroma cacao) Germplasm and Development of Molecular Methods for Use in Diagnostics. *Pathogens*, *12*(11), 1284. https://doi.org/10.3390/pathogens12111284

Umber, M., Pressat, G., Fort, G., Plaisir Pineau, K., Guiougiou, C., Lambert, F., Farinas, B., Pichaut, J. P., Janzac, B., Delos, J. M., Salmon, F., Dubois, C., & Teycheney, P. Y. (2022). Risk Assessment of Infectious Endogenous Banana Streak Viruses in Guadeloupe. *Frontiers in Plant Science*, *13*. https://doi.org/10.3389/fpls.2022.951285

Untung, K. (2000). Pelembagaan Konsep Pengendalian Hama Terpadu di Indonesia. *Jurnal Perlindungan Tanaman Indonesia*, *6*(1), 1–8.

Veglia, A. J., Bistolas, K. S. I., Voolstra, C. R., Hume, B. C. C., Ruscheweyh, H. J., Planes, S., Allemand, D., Boissin, E., Wincker, P., Poulain, J., Moulin, C., Bourdin, G., Iwankow, G., Romac, S., Agostini, S., Banaigs, B., Boss, E., Bowler, C., de Vargas, C., Vega Thurber, R. L. (2023). Endogenous viral elements reveal associations between a non-retroviral RNA virus and symbiotic dinoflagellate genomes. *Communications Biology*, *6*(1). https://doi.org/10.1038/s42003-023-04917-9

Voora, V., Bermudez, S., & Larrea, C. (2019). *Global Market Report : Cocoa*. https://iisd.org/ssi/. Diakses pada 02 Januari 2021

Waterworth, R. A., Wright, I. M., & Millar, J. G. (2011). Reproductive Biology of Three Cosmopolitan Mealybug (Hemiptera: Pseudococcidae) Species, Pseudococcus

longispinus, Pseudococcus viburni, and Planococcus ficus. *Ann. Entomol. Soc. Am*, *104*(2), 249–260. https://doi.org/10.1603/AN10139

Yu, H., Wang, X., Lu, Z., Xu, Y., Deng, X., & Xu, Q. (2019). Endogenous pararetrovirus sequences are widely present in Citrinae genomes. *Virus Research*, *262*, 48–53. https://doi.org/10.1016/j.virusres.2018.05.018

Zarkani, A., Apriyanto, D., Turanli, F., Ercan, C., & Kaydan, M. B. (2021). A checklist of Indonesian scale insects (Hemiptera: Coccomorpha). *Zootaxa*, *5016*(2), 151–195. https://doi.org/10.11646/zootaxa.5016.2.1

Zarkani, A., Fauzi, A., Apriyanto, D., & Kaydan, M. B. (2023). Mealybugs (Hemiptera, Coccomorpha, Pseudococcidae) on parasitic plants (Loranthaceae) in Indonesia with description of a new species and a new country record. *ZooKeys*, *1167*, 199–210. https://doi.org/10.3897/zookeys.1167.106012