

DAFTAR PUSTAKA

- Agostini-Costa, D. T. S., Vieira, R. F., Bizzo, H. R., Silveira, D., and Gimenes, M. A. (2012). Secondary Metabolites. Brazil.
- Ahmad, A., Ali, H., Khan, H., Begam, A., Khan, S., Ali, S. S., Ahmad, N., Fazal, H., Ali, M., Hano, C., Ahmad, N., and Abbasi, B. H. (2020). Effect of gibberellic acid on production of biomass, polyphenolics and steviol glycosides in adventitious root cultures of *Stevia rebaudiana* (Bert.). *Plants*, 9(4). <https://doi.org/10.3390/plants9040420>.
- Al-Sane', K. O., Shibli, R. A., Freihat, N. M., and Hammouri, M. K. (2005). Cell Suspension Culture and Secondary Metabolites Production in African Violet (*Saintpaulia ionantha* Wendl.). *Jordan Journal of Agricultural Sciences* (Vol. 1, Issue 1).
- Anggraito, Y. U., Susanti, R., Iswari, R. S., Yuniastuti, A., Lisdiana, WH, N., Habibah, N. A., and Bintari, S. H. (2018). Metabolit Sekunder dari Tanaman: Aplikasi dan Produksi. Fakultas Matematika dan Ilmu Pengetahuan Alam Universitas Negeri Semarang.
- Anju, T., Rai, N. K. S. R., and Kumar, A. (2022). *Sauropus androgynus* (L.) Merr. A Multipurpose Plant with Multiple Uses in Traditional Ethnic Culinary and Ethnomedicinal Preparations. *Journal of Ethnic Foods* (Vol. 9, Issue 1). BioMed Central Ltd. <https://doi.org/10.1186/s42779-022-00125-8>.
- AOAC. 2005. Official Methods of Analysis of AOAC International. 18th Ed. Assoc. Off. Anal. Chem., Arlington.
- Arif, T. (2020). Therapeutic potential and traditional uses of *Sauropus androgynous*: A review Tamanna Arif and Raviraja Shetty G. ~ 2131 ~ *Journal of Pharmacognosy and Phytochemistry*, 9(3). www.phytojournal.com.
- Asra, R. (2014). Pengaruh Hormon Giberelin (GA₃) Terhadap Daya Kecambah dan Vigoritas *Calopogonium caeruleum*. *Biospecies* Vol. 7 No.1, Januari 2014, hal. 29-33.
- Asra, R., dan Ubaidillah, D. (2012). Pengaruh Konsentrasi Giberelin (GA₃) Terhadap Nilai Nutrisi *Calopogonium caeruleum*. *Jurnal Ilmu-Ilmu Peternakan* (Issue 2).
- Azeez, H. A., and Ibrahim, K. M. (2014). Hypericum Triquetrifolium Callus Cultures a Potential Source of Phenolics and Flavonoids. *Journal of Zankoy Sulaimani-Part A, Special Issue* (Vol. 16).
- Bačėninaitė, D., Džermeikaitė, K., and Antanaitis, R. (2022). Global Warming and Dairy Cattle: How to Control and Reduce Methane Emission. *Animals* (Vol. 12, Issue 19). MDPI. <https://doi.org/10.3390/ani12192687>.

- Bai, W. Q., Xiao, Y. H., Zhao, J., Song, S. Q., Hu, L., Zeng, J. Y., Li, X. B., Hou, L., Luo, M., Li, D. M., and Pei, Y. (2014). Gibberellin overproduction promotes sucrose synthase expression and secondary cell wall deposition in cotton fibers. *PLoS ONE*, 9(5). <https://doi.org/10.1371/journal.pone.0096537>.
- Ballhorn, D. J., Kautz, S., Heil, M., and Hegeman, A. D. (2009). Cyanogenesis of wild lima bean (*Phaseolus lunatus* L.) is an efficient direct defence in nature. *PLoS ONE*, 4(5). <https://doi.org/10.1371/journal.pone.0005450>.
- Cahyani, R. D., Nuswantara, L. K., dan Subrata, D. A. (2012). Pengaruh Proteksi Protein Tepung Kedelai dengan Tanin Daun Bakau Terhadap Konsentrasi Amonia, Undegraded Protein dan Protein Total Secara. *Animal Agricultural Journal*, 1(1), 159–166. <http://ejournal-s1.undip.ac.id/index.php/aaaj>.
- Cardoso-Gutierrez, E., Aranda-Aguirre, E., Robles-Jimenez, L. E., Castelán-Ortega, O. A., Chay-Canul, A. J., Foggi, G., Angeles-Hernandez, J. C., Vargas-Bello-Pérez, E., and González-Ronquillo, M. (2021). Effect of tannins from tropical plants on methane production from ruminants: A systematic review. In *Veterinary and Animal Science* (Vol. 14). Elsevier B.V. <https://doi.org/10.1016/j.vas.2021.100214>.
- Castro-Camba, R., Sánchez, C., Vidal, N., and Vielba, J. M. (2022). Plant Development and Crop Yield: The Role of Gibberellins. *Plants* (Vol. 11, Issue 19). MDPI. <https://doi.org/10.3390/plants11192650>.
- Chatterjee, S., Mukherjee, D., Mani, A., and Choudhuri, P. (2019). Nutritional Status of Chekurmanis (*Sauropus androgynous*): Vol. (I. Chakraborty, Riadhllahy, B. Vikram, S. O.J, and A. Mani, Eds.). Satish Serial Publishing House.
- Daie A', J., Watts, M., Aloni B', B., and Wyse B', R. E. (1986). In Vitro and In Vivo Modification of Sugar Transport and Translocation in Celery by Phytohormones. *Plant Science* (Vol. 46).
- Davière, J. M., and Achard, P. (2016). A Pivotal Role of DELLAs in Regulating Multiple Hormone Signals. *Molecular Plant* (Vol. 9, Issue 1, pp. 10–20). Cell Press. <https://doi.org/10.1016/j.molp.2015.09.011>.
- Davies, P. J. (1995). A. The Plant Hormones: Their Nature, Occurrence, and Functions.
- Dewi, N. P. (2020). Uji Kualitatif dan Kuantitatif Metabolit Sekunder Ekstrak Etanol Daun Awar-Awar (*Ficus septica* Burm. f) dengan Metode Spektrofotometer UV-VIS. *Acta Holist. Pharm*, 2(1), 16–24.
- Dewick, P. M. (2002). Medicinal natural products: a biosynthetic approach (2nd ed.). John Wiley and Sons Ltd.

- Dong, Y., Guo, W., Xiao, W., Liu, J., Jia, Z., Zhao, X., Jiang, Z., and Chang, E. (2023). Effects of Different Donor Ages on the Growth of Cutting Seedlings Propagated from Ancient *Platycladus orientalis*. *Plants*, 12(9). <https://doi.org/10.3390/plants12091754>.
- Eilert, U., De Luca, V., Constabel, F., and Kurz, W. G. W. (1987). Elicitor-Mediated Induction of Tryptophan Decarboxylase and Strictosidine Synthase Activities in Cell Suspension Cultures of *Catharanthus roseus*. In *Archives of Biochemistry and Biophysics* (Vol. 254, Issue 2).
- Engel, A. M., Klevenhusen, F., Moenning, J. L., Numata, J., Fischer-Tenhagen, C., Sachse, B., Schäfer, B., Fry, H., Kappenstein, O., and Pieper, R. (2022). Investigations on the Transfer of Quinolizidine Alkaloids from *Lupinus angustifolius* into the Milk of Dairy Cows. *Journal of Agricultural and Food Chemistry*, 70(37), 11749–11758. <https://doi.org/10.1021/acs.jafc.2c02517>.
- Enny, M., and Lidar, S. (2018). Respon Tanaman Pakcoy (*Brassica rapa* L) Akibat Pemberian Zat Pengatur Tumbuh Hormonik. *Jurnal Ilmiah Pertanian*, Vol. 14(No.2).
- Facchini, P. J. (2001). Alkaloid Biosynthesis in Plants: Biochemistry, Cell Biology, Molecular Regulation, and Metabolic Engineering Applications. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* (Vol. 52). www.annualreviews.org.
- Fathonah, D., and Sugiyarto, S. (2019). Effect of IAA and GA₃ Toward the Growing and Saponin Content of Purwaceng (*Pimpinella alpina*). *Nusantara Bioscience*, 1. <https://doi.org/10.13057/nusbiosci/n010103>.
- Fonouni-Farde, C., Miassod, A., Laffont, C., Morin, H., Bendahmane, A., Diet, A., and Frugier, F. (2019). Gibberellins negatively regulate the development of *Medicago truncatula* root system. *Scientific Reports*, 9(1). <https://doi.org/10.1038/s41598-019-38876-1>.
- Gagné, S., Lacampagne, S., Claisse, O., and Gény, L. (2009). Leucoanthocyanidin Reductase and Anthocyanidin Reductase Gene Expression and Activity In Flowers, Young Berries and Skins of *Vitis Vinifera* L. Cv. Cabernet-Sauvignon during Development. *Plant Physiology and Biochemistry*, 47 (4), 282–290. <https://doi.org/10.1016/j.plaphy.2008.12.004>.
- Geerlings, A., Martinez-Lozano Ibañez, M., Memelink, J., Van Der Heijden, R., and Verpoorte, R. (2000). Molecular cloning and analysis of strictosidine β-D-glucosidase, an enzyme in terpenoid indole alkaloid biosynthesis in *Catharanthus roseus*. *Journal of Biological Chemistry*, 275(5), 3051–3056. <https://doi.org/10.1074/jbc.275.5.3051>.
- Handayani, S., Setyawati, I., Ariendha, D. S. R., Pratiwi, Y. S., Idyawati, S., and Fatmawati, N. (2020). The Effect of Katuk Leaf (*Sauropus androgynus* L. Merr.) Biscuit Consumption toward Increasing Breastmilk

Volume on the 10th Day. Journal of Physics: Conference Series, 1594(1). <https://doi.org/10.1088/1742-6596/1594/1/012051>.

Hanum, C. (2013). Pertumbuhan, Hasil, dan Mutu Biji Kedelai dengan Pemberian Pupuk Organik dan Fosfor. *J. Agron. Indonesia*, 41(3), 209–214.

Harberd, N. P., Belfield, E., and Yasumura, Y. (2009). The Angiosperm Gibberellin-GID1-DELLA Growth Regulatory Mechanism: How an “Inhibitor of an Inhibitor” Enables Flexible Response to Fluctuating Environments. *Plant Cell* (Vol. 21, Issue 5, pp. 1328–1339). American Society of Plant Biologists. <https://doi.org/10.1105/tpc.109.066969>.

Haslam, E. (2007). Vegetable Tannins - Lessons of a Phytochemical Lifetime. *Phytochemistry* (Vol. 68, Issues 22–24, pp. 2713–2721). <https://doi.org/10.1016/j.phytochem.2007.09.009>.

Hassanein, A. M. A. (2013). Factors Influencing Plant Propagation Efficiency Via Stem Cuttings. *Journal of Horticultural Science and Ornamental Plants*, 5(3), 171–176. <https://doi.org/10.5829/idosi.jhsop.2013.5.3.1125>.

Heath, R., White, S., and Rock, C. (2002). Inhibitors of Fatty Acid Synthesis as Antimicrobial Chemotherapeutics. *Applied Microbiology and Biotechnology* (Vol. 58, Issue 6, pp. 695–703). <https://doi.org/10.1007/s00253-001-0918-z>.

Heldt, H.-W., Piechulla, B., and Heldt, F. (2011). *Plant Biochemistry* Fourth edition (4th ed.). Academic Press is an imprint of Elsevier. Department in Oxford, United Kingdom.

Hidayanto, M., Nurjanah, S., dan Yossita, F (2003). Pengaruh Panjang Stek Akar dan Konsentrasi Natrium-Nitrofenol Terhadap Pertumbuhan Stek Akar Sukun (*Artocarpus communis* F.). *Jurnal Pengkajian dan Pengembangan Teknologi Pertanian* (Vol. 6, Issue 2).

Hoe, V. B., and Siong, K. H. (1999). The Nutritional of Indigenous Fruits and Vegetables in Sarawak. *Asia Pacific J Clin Nitr*, 8(1), 24–31. <https://doi.org/https://doi.org/10.1046/J.1440-6047.1999.00046.X>.

Horseman, N. D., and Gregerson, K. A. (2015). Prolactin. In *Endocrinology: Adult and Pediatric* (Vols. 1–2, pp. 91-103.e4). Elsevier Inc. <https://doi.org/10.1016/B978-0-323-18907-1.00006-8>.

Inoue, S., Ilogu, C., and Sobze, J.-M. (2023). Effects of Indole-3-Butyric Acid and Age of Stem Cuttings on Root Morphology, Growth, and Survival of *Cornus Sericea*. *Journal of Forestry Research*, 34(2), 433–440. <https://doi.org/10.1007/s11676-022-01490-5>.

Isah, T. (2019). Stress and Defense Responses in Plant Secondary Metabolites Production. *Biological research* (Vol. 52, Issue 1, p. 39). *NLM (Medline)*. <https://doi.org/10.1186/s40659-019-0246-3>.

- Jamwal, K., Bhattacharya, S., and Puri, S. (2018a). Plant Growth Regulator Mediated Consequences of Secondary Metabolites in Medicinal Plants. *Journal of Applied Research on Medicinal and Aromatic Plants* (Vol. 9, pp. 26–38). Elsevier GmbH. <https://doi.org/10.1016/j.jarmap.2017.12.003>.
- Jamwal, K., Bhattacharya, S., and Puri, S. (2018b). Plant Growth Regulator Mediated Consequences of Secondary Metabolites in Medicinal Plants. *Journal of Applied Research on Medicinal and Aromatic Plants* (Vol. 9, pp. 26–38). Elsevier GmbH. <https://doi.org/10.1016/j.jarmap.2017.12.003>.
- Jerónimo, E., Lamy, E. C., Pinheiro, C. M. S. C., and Dentinho, M. (2016). *Tannins in Ruminant Nutrition: Impact on Animal Performance and Quality of Edible Products*. Nova Science Publishers New York.
- Jim, G. 2015. Katuk (*Sauropus androgynus*). <https://growerjim.blogspot.com/2015/01/katuk-sauropus-androgynus>. Diakses pada 19 Januari 2024.
- Julianto, T. S. (2019). *Fitokimia Tinjauan Metabolit Sekunder dan Skrining Fitokimia* (1st ed.). Universitas Islam Indonesia.
- Katyayini, N. U., Rinne, P. L. H., Tarkowská, D., Strnad, M., and van der Schoot, C. (2020). Dual Role of Gibberellin in Perennial Shoot Branching: Inhibition and Activation. *Frontiers in Plant Science*, 11. <https://doi.org/10.3389/fpls.2020.00736>.
- Kementerian Kesehatan Republik Indonesia. (2017). *Farmakope Herbal Indonesia (II)*. Kementerian Kesehatan Republik Indonesia.
- Khalid, A., and Aftab, F. (2020). Effect of exogenous application of IAA and GA₃ on growth, protein content, and antioxidant enzymes of *Solanum tuberosum* L. grown in vitro under salt stress. *In Vitro Cellular and Developmental Biology - Plant*, 56(3), 377–389. <https://doi.org/10.1007/s11627-019-10047-x>.
- Khristyana, L., Anggarwulan, E., and Marsusi, M. (2005). Growth, Saponin and Nitrogen Content of Common Plantain (*Plantago major* L.) Tissue with Gibberellic Acid Application (GA₃). *Biofarmasi Journal of Natural Product Biochemistry*, 3(1), 11–15. <https://doi.org/10.13057/biofar/f030103>.
- Kim, H. J., and Triplett, B. A. (2001). Cotton Fiber Growth in Planta And in Vitro. Models For Plant Cell Elongation and Cell Wall Biogenesis. *Plant Physiology*, 127(4), 1361–1366. <https://doi.org/10.1104/pp.010724>.
- Koester, L. R., Pool, D. H., Serão, N. V. L., and Schmitz-Esser, S. (2020). Beef cattle that respond differently to fescue toxicosis have distinct gastrointestinal tract microbiota. *PLoS ONE*, 15(7). <https://doi.org/10.1371/journal.pone.0229192>.

- Laveena, K. B., and Chandra, M. (2018). Evaluation of Bioactive Compounds, Antioxidant, and Antibacterial Properties of Medicinal Plants *Sauropus Androgynus* L. and *Erythrina variegata* L. Asian Journal of Pharmaceutical and Clinical Research, 11(12), 313–317. <https://doi.org/10.22159/ajpcr.2018.v11i12.28207>.
- Liu, H., Vaddella, V., and Zhou, D. (2011). Effects of Chestnut Tannins and Coconut Oil on Growth Performance, Methane Emission, Ruminal Fermentation, and Microbial Populations in Sheep. Journal of Dairy Science, 94(12), 6069–6077. <https://doi.org/10.3168/jds.2011-4508>.
- Locascio, A., Blázquez, M. A., and Alabadí, D. (2013). Dynamic Regulation of Cortical Microtubule Organization Through Prefoldin-DELLA interaction. Current Biology, 23(9), 804–809. <https://doi.org/10.1016/j.cub.2013.03.053>.
- Maisuthisakul, P., Pasuk, S., and Ritthiruangdej, P. (2008). Relationship Between Antioxidant Properties and Chemical Composition of Some Thai Plants. Journal of Food Composition and Analysis, 21(3), 229–240. <https://doi.org/10.1016/j.jfca.2007.11.005>.
- Mannelli, F., Daghighi, M., Alves, S. P., Bessa, R. J. B., Minieri, S., Giovannetti, L., Conte, G., Mele, M., Messina, A., Rapaccini, S., Viti, C., and Buccioni, A. (2019). Effects of Chestnut Tannin Extract, Vescalagin and Gallic Acid on the Dimethyl Acetals Profile and Microbial Community Composition in Rumen Liquor: An *In Vitro* Study. Microorganisms, 7(7). <https://doi.org/10.3390/microorganisms7070202>.
- Maryati, Y., Susilowati, A., Aspiyanto, Mulyani, H., Artanti, N., and Budiari, S. (2019). Evaluation of Antioxidant Activity of Formulated Functional Drinks Derived from Katuk (*Sauropus androgynus*) Leaf Extracts: Optimization Using Response Surface Methodology (RSM). AIP Conference Proceedings, 2175. <https://doi.org/10.1063/1.5134582>.
- Mastiningsih, N. P. (2021). The Impact of Moringa Leaves, Katuk Leaves and Oxytocin Massage on Quantity and Quality of Mother's Milk as Patients Health Center in Bali. Journal of Environmental Treatment Techniques, 9(1), 275–279. [https://doi.org/10.47277/jett/9\(1\)279](https://doi.org/10.47277/jett/9(1)279).
- Matos, F. S., Freitas, I. A. S., Pereira, V. L. G., and Pires, W. K. L. (2020). Effect of Gibberellin on Growth and Development of *Spondias tuberosa* Seedlings. Revista Caatinga, 33(4), 1124–1130. <https://doi.org/10.1590/1983-21252020v33n427rc>.
- Mayerni, R. (2008). Pengaruh Beberapa Konsentrasi Giberelin Terhadap Pertumbuhan Bibit Kina Succi (*Cinchona succirubra* Pavon). Jerami Indonesian Journal of Crop Science, 1(1).
- Melchior, E. A., and Myer, P. R. (2018). Fescue toxicosis and Its Influence on the Rumen Microbiome: Mitigation of Production Losses Through Clover

Isoflavones. *Journal of Applied Animal Research*, 46(1), 1280–1288.
<https://doi.org/10.1080/09712119.2018.1496920>.

Menke, F. L. H., Parchmann, S., Mueller, M. J., Kijne, J. W., and Memelink, J. (1999). Involvement of the Octadecanoid Pathway and Protein Phosphorylation in Fungal Elicitor-Induced Expression of Terpenoid Indole Alkaloid Biosynthetic Genes in *Catharanthus roseus*. www.plantphysiol.org.

Miceli, A., Moncada, A., Sabatino, L., and Vetrano, F. (2019). Effect of Gibberellic Acid on Growth, Yield, and Quality of Leaf Lettuce and Rocket Grown in A Floating System. *Agronomy*, 9(7).
<https://doi.org/10.3390/agronomy9070382>.

Michels, A., Neumann, M., Leão, G. F. M., Reck, A. M., Bertagnon, H. G., Lopes, L. S., De Souza, A. M., Dos Santos, L. C., and Júnior, E. S. S. (2018). Isoquinoline Alkaloids Supplementation on Performance And Carcass Traits of Feedlot Bulls. *Asian-Australasian Journal of Animal Sciences*, 31(9), 1474–1480. <https://doi.org/10.5713/ajas.17.0868>.

Miean, K. H., and Mohamed, S. (2001). Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) Content of Edible Tropical Plants. *Journal of Agricultural and Food Chemistry*, 49(6), 3106–3112.
<https://doi.org/10.1021/jf000892m>.

Missouri Botanical Garden, 2024. *Sauropus androgynus* (L.) Merr.
<https://www.tropicos.org/name/12800096>. Diakses pada 19 Januari 2024.

Mitchum, M. G., Yamaguchi, S., Hanada, A., Kuwahara, A., Yoshioka, Y., Kato, T., Tabata, S., Kamiya, Y., and Sun, T. P. (2006). Distinct and Overlapping Roles of two Gibberellin 3-oxidases in Arabidopsis Development. *Plant Journal*, 45(5), 804–818.
<https://doi.org/10.1111/j.1365-313X.2005.02642.x>.

Mole, S., Rogler, J. C., and Butlert, L. G. (1993). Growth Reduction by Dietary Tannins: Different Effects Due to Different Tannins. *Biochemical Systematics and Ecology* (Vol. 21, Issue 6).

Mollah, A. F., Rafiq, Z., Saiful, M., Sarker, A., Nazmul, M., and Rony, H. (2019). Effect of Plant Age for Cuttings on The Growth and Seed Yield in Late Season Jute Under Different Planting Spacings.
<https://www.researchgate.net/publication/337020696>.

Mostafa, G. G., and Alhamd, M. F. Abou. (2011). Effect of Gibberellic acid and Indole 3-acetic acid on Improving Growth and Accumulation of Phytochemical Composition in *Balanites aegyptiaca* Plants. *American Journal of Plant Physiology*, 6(1), 36–43.
<https://doi.org/10.3923/ajpp.2011.36.43>.

- Mueller, M. J., Brodschelm, W., Spannagl, E., and Zenk, M. H. (1993). Signaling in The Elicitation Process is Mediated Through The Octadecanoid Pathway Leading To Jasmonic Acid. *Proc. Natl. Acad. Sci. USA* (Vol. 90).
- Naumann, H. D., Tedeschi, L. O., Muir, J. P., Lambert, B. D., and Kothmann, M. M. (2013). Effect of Molecular Weight of Condensed Tannins from Warm-Season Perennial Legumes on Ruminal Methane Production Invitro. *Biochemical Systematics and Ecology*, 50, 154–162. <https://doi.org/10.1016/j.bse.2013.03.050>.
- Naumann, H. D., Tedeschi, L. O., Zeller, W. E., and Huntley, N. F. (2017). The role of condensed tannins in ruminant animal production: Advances, limitations and future directions. *Revista Brasileira de Zootecnia* (Vol. 46, Issue 12, pp. 929–949). Sociedade Brasileira de Zootecnia. <https://doi.org/10.1590/S1806-92902017001200009>.
- Naveena, E., Janavi, G., Arumugam, T., and Anitha, T. (2020). Estimation of Nutritive Composition of *Sauropus androgynus* (Multivitamin plant) at Different Growth Stages and Position of Leaves. *International Journal of Chemical Studies*, 8(3), 443–447. <https://doi.org/10.22271/chemi.2020.v8.i3e.9251>.
- Nishi, A., Kuroiwa, M., Miller, D. B., O'Callaghan, J. P., Bateup, H. S., Shuto, T., Sotogaku, N., Fukuda, T., Heintz, N., Greengard, P., and Snyder, G. L. (2008). Distinct Roles of PDE4 and PDE10A in the Regulation of cAMP/PKA Signaling in the Striatum. *Journal of Neuroscience*, 28(42), 10460–10471. <https://doi.org/10.1523/JNEUROSCI.2518-08.2008>
- Padmavathi, P., and Rao, and M. P. (1990). Nutritive Value of *Sauropus androgynus* leaves*. *Plant Foods for Human Nutrition* (Vol. 40).
- Pandey, R., Paul, V Das. M, Meena, M And Meena. RC. 2017. Plant Growth Analysis. <https://doi.org/10.13140/RG.2.2.21657.72808>.
- Pasquali, G., Goddijn, O. J., de Waal, A., Verpoorte, R., Schilperoort, R. A., Hoge, H. C., and Memelink, J. (1992). Coordinated Regulation of Two Indole Alkaloid Biosynthetic Genes from *Catharanthus Roseus* by Auxin and Elicitors. *Plant Molecular Biology* (Vol. 18).
- Pavlista, A. D., Santra, D. K., Schild, J. A., and Hergert, G. W. (2012). Gibberellic Acid Sensitivity Among Common Bean Cultivars (*Phaseolus vulgaris* L.). *HortScience*, 47(5), 637–642. <https://doi.org/10.21273/hortsci.47.5.637>.
- Petrus, A. J. A. (2013). *Sauropus androgynus* (L.) merrill A Potentially Nutritive Functional Leafy-Vegetable. *Asian Journal of Chemistry* (Vol. 25, Issue 17, pp. 9425–9433). Chemical Publishing Co. <https://doi.org/10.14233/ajchem.2013.15405>.

- Pertiwi, P. D, Agustiansyah, Nurmiaty, Y. (2014). Pengaruh Giberelin (GA3) Terhadap Pertumbuhan dan Produksi Tanaman Kedelai (*Glycine max* (L.) Merrill.). Jurnal Agrotek Tropika (Vol. 2, Issue 2).
- Platel, K., and Srinivasan, K. (2017). Nutritional Profile of Chekurmanis (*Sauropus androgynus*), A Less Explored Green Leafy Vegetable. The Indian Journal of Nutrition and Dietetics, 54(3), 243. <https://doi.org/10.21048/ijnd.2017.54.3.15765>.
- Purba, R. A. P., and Paengkoum, P. (2022). Exploring the Phytochemical Profiles and Antioxidant, Antidiabetic, and Antihemolytic Properties of *Sauropus androgynus* Dried Leaf Extracts for Ruminant Health and Production. *Molecules*, 27(23). <https://doi.org/10.3390/molecules27238580>.
- Quan, J., Ni, R., Wang, Y., Sun, J., Ma, M., and Bi, H. (2022). Effects of Different Growth Regulators on the Rooting of *Catalpa bignonioides* Softwood Cuttings. *Life*, 12(8). <https://doi.org/10.3390/life12081231>.
- Raidas, D., Krishi, S., Pradesh, M., Upadhayaya, I. S., Sharma, I. A., Choudhary, I. A., Upadhayaya, S., Sharma, A., and Choudhary, A. (2019). Impact of Plant Growth Hormones on Growth, Physiology and Alkaloid Content of Kalmegh (*Andrographis peniculata* Burn F. Ex). *Journal of Pharmacognosy and Phytochemistry*, 2, 715–718. <https://www.phytojournal.com/archives/2019/vol8issue2S/PartS/SP-8-2-193-104.pdf>.
- Rao, S. R., and Ravishankar, G. A. (2002). Plant cell cultures: Chemical factories of secondary metabolites. [https://doi.org/https://doi.org/10.1016/S0734-9750\(02\)00007-1](https://doi.org/https://doi.org/10.1016/S0734-9750(02)00007-1).
- Rathbone, D. A., and Bruce, N. C. (2002). Microbial transformation of alkaloids. [https://doi.org/https://doi.org/10.1016/S1369-5274\(02\)00317-X](https://doi.org/https://doi.org/10.1016/S1369-5274(02)00317-X).
- Ritonga, F. N., Zhou, D., Zhang, Y., Song, R., Li, C., Li, J., and Gao, J. (2023). The Roles of Gibberellins in Regulating Leaf Development. *Plants* (Vol. 12, Issue 6). MDPI. <https://doi.org/10.3390/plants12061243>.
- Rohmawati, I. (2013). Penentuan Dosis Pemupukan N, P dan K Pada Budidaya Katuk (*Sauropus androgynus* (L.) Merr.). Tesis. Sekolah Pascasarjana Institut Pertanian Bogor. Bogor. <https://adoc.pub/penentuan-dosis-pemupukan-n-p-dan-k-pada-budidaya-katuk-saur.html>.
- Saidi, I. A., Azara, R., dan Yanti, E. (2021). Buku Ajar Pasca Panen dan Pengolahan Sayuran Daun Penulis (M. Tanzil Multazam and Mahardika Darmawan, Eds.; 1st ed.). Umsida Press.
- Santana, T., Rahayu, A., dan Mulyaningsih Mahasiswa, Y. S. (2021). Karakterisasi Morfologi dan Kualitas Berbagai Aksesori Katuk (*Sauropus*

androgynus (L.) Merr.). Jurnal Agronida ISSN (Vol. 7, Issue 1).
<https://doi.org/https://doi.org/10.30997/jag.v7i1.4102>.

Santoso, I. U. (2013). Katuk, Tumbuhan Multi Khasiat: Vol. (R. Saepudin, Ed.). Fakultas Pertanian (BPFP) Unib. <https://www.researchgate.net/publication/303994522>.

Schrader, J., Shi, P., Royer, D. L., Peppe, D. J., Gallagher, R. V., Li, Y., Wang, R., and Wright, I. J. (2021). Leaf Size Estimation Based on Leaf Length, Width and Shape. *Annals of Botany*, 128(4), 395–406.
<https://doi.org/10.1093/aob/mcab078>.

Singer, S. D., Zou, J., and Weselake, R. J. (2016). Abiotic factors influence plant storage lipid accumulation and composition. *Plant Science* (Vol. 243, pp. 1–9). Elsevier Ireland Ltd.
<https://doi.org/10.1016/j.plantsci.2015.11.003>.

Singh, S., Singh, D. R., Salim, K. M., Srivastava, A., Singh, L. B., and Srivastava, R. C. (2011a). Estimation of Proximate Composition, Micronutrients and Phytochemical Compounds in Traditional Vegetables from Andaman and Nicobar Islands. *International Journal of Food Sciences and Nutrition*, 62(7), 765–773.
<https://doi.org/10.3109/09637486.2011.585961>.

Singh, S., Singh, D. R., Salim, K. M., Srivastava, A., Singh, L. B., and Srivastava, R. C. (2011b). Estimation of Proximate Composition, Micronutrients and Phytochemical Compounds in Traditional Vegetables from Andaman and Nicobar Islands. *International Journal of Food Sciences and Nutrition*, 62(7), 765–773.
<https://doi.org/10.3109/09637486.2011.585961>.

Siregar, N. (2014). Pengaruh Umur Bahan Setek Terhadap Pertumbuhan Setek Akor (*Acacia auriculiformis* A. Cunn. Ex Benth). *Jurnal Perbenihan Tanaman Hutan*, 2(2), 109–117.

Soka, S., Alam, H., Boenjamin, N., Agustina, T. W., and Suhartono, M. T. (2010). Effect of *Sauropus androgynus* Leaf Extracts on The Expression of Prolactin and Oxytocin Genes in Lactating BALB/C Mice. *Journal of Nutrigenetics and Nutrigenomics*, 3(1), 31–36.
<https://doi.org/10.1159/000319710>.

Sousa, M. C., Bronzatto, A. C., González-Esquinca, A. R., Campos, F. G., Dalanhol, S. J., Boaro, C. S. F., Martins, A. L., da Silva Almeida, J. R. G., Costa, E. V., De-la-Cruz-Chacón, I., and Ferreira, G. (2019). The Production of Alkaloids in *Annona* Seedlings is Affected by the application of GA4+7 + 6-Benzyladenine. *Biochemical Systematics and Ecology*, 84, 47–51. <https://doi.org/10.1016/j.bse.2019.03.007>.

Sunardi, O., Adimihardja, S. A., and Mulyaningsih, Y. (2013). Effect of Giving Gibberellin (GA3) on Vegetative Plant Growth Water Cabbage (*Ipomea*

Aquatica Forsk L.) in The Floating Raft Technique (FRT) Hydroponic System. In *Jurnal Pertanian ISSN 2087* (Vol. 4936).

Suprpto, A. (2004). Auksin: Zat Peengatur Tumbuh Penting Meningkatkan Mutu Stek Tanaman. *Jurnal Penelitian Inovasi*, 21(no 1), 81–90. <https://www.neliti.com/publications/17658/>.

Suprayogi, A., Kusumorini, N., dan Evi Dame Arita, S. (2015). Fraksi Heksan Daun Katuk Sebagai Obat Untuk Memperbaiki Produksi Susu, Penampilan Induk, dan Anak Tikus. *Jurnal Veteriner*, 16(N0. 1), 88–95.

Suprayogi, A., Latif, H., dan Yayan Ruhyana, A. (2013). Peningkatan Produksi Susu Sapi Perah di Peternakan Rakyat Melalui Pemberian Katuk-IPB3 sebagai Aditif Pakan (Increasing Milk Production of Dairy Cattle in the Farm by Giving Katuk-IPB3 as Feed Additive). *Jurnal Ilmu Pertanian Indonesia (JIPI)*, Desember (Vol. 18, Issue 3). <http://www.surabayapagi.com>.

Susanti, N. M. P., Budiman, I. N. A., dan Warditiani, N. K. (2014). Skrining Fitokimia Ekstrak Etanol 90% Daun Katuk (*Sauropus androgynus* (L.) Merr.). *Jurnal Farmasi Udayana*, 3(No 1), 83–86. <https://www.neliti.com/publications/279778/>.

Syamsiah, M., Pd, S., Si, M., Marlina, G., dan Agr, S. (2016). Respon Pertumbuhan Tanaman Selada (*Lactuca sativa* L.) Varietas Krieba Terhadap Konsentrasi Asam Giberelin Melissa Syamsiah dan Gina Marlina 55. In *Journal of Agrosience* (Vol. 6, Issue 2).

Tedeschi, L. O., Muir, J. P., Naumann, H. D., Norris, A. B., Ramírez-Restrepo, C. A., and Mertens-Talcott, S. U. (2021). Nutritional Aspects of Ecologically Relevant Phytochemicals in Ruminant Production. In *Frontiers in Veterinary Science* (Vol. 8). Frontiers Media S.A. <https://doi.org/10.3389/fvets.2021.628445>.

Utami. (2018). Pengaruh Hormon Tumbuh Terhadap Fisiologi Tanaman (Suatu Kajian Pustaka). Prodi Agroekoteknologi Fakultas Pertanian Universitas Udayana.

Waghorn, G. (2008). Beneficial and Detrimental Effects of Dietary Condensed Tannins for Sustainable Sheep and goat production-Progress and Challenges. *Animal Feed Science and Technology*, 147(1–3), 116–139. <https://doi.org/10.1016/j.anifeedsci.2007.09.013>.

Waghorn, G. C., and McNabb, W. C. (2003). Consequences of Plant Phenolic Compounds for Productivity and Health of Ruminants. *Proceedings of the Nutrition Society*, 62(2), 383–392. <https://doi.org/10.1079/pns2003245>.

War, A. R., Paulraj, M. G., Ahmad, T., Buhroo, A. A., Hussain, B., Ignacimuthu, S., and Sharma, H. C. (2012). Mechanisms of Plant

Defense Against Insect Herbivores. Plant Signaling and Behavior (Vol. 7, Issue 10). Landes Bioscience. <https://doi.org/10.4161/psb.21663>.

Wiraatmaja. (2017). Zat Pengatur Tumbuh Giberelin dan Sitokinin. Program Studi Agroekoteknologi, Fakultas Pertanian, Udayana.

Wu, G. (2018). Principles of Animal Nutrition. CRC Press Taylor and Francis Group. <http://www.crcpress.com>.

Wuyts, N., De Waele, D., and Swennen, R. (2006). Extraction And Partial Characterization of Polyphenol Oxidase from Banana (*Musa Acuminata* Grande Naine) Roots. Plant Physiology and Biochemistry, 44(5–6), 308–314. <https://doi.org/10.1016/j.plaphy.2006.06.005>.

Xia, Q., Wu, W. C., Tian, K., Jia, Y. Y., Wu, X., Guan, Z., and Tian, X. J. (2015). Effects of Different Cutting Traits on Bud Emergence and Early Growth of the Chinese Vegetable Toona Sinensis. Scientia Horticulturae, 190, 137–143. <https://doi.org/10.1016/j.scienta.2015.04.026>.

Xiao, G., Zhao, P., and Zhang, Y. (2019). A Pivotal Role of Hormones in Regulating Cotton Fiber Development. In Frontiers in Plant Science (Vol. 10). Frontiers Media S.A. <https://doi.org/10.3389/fpls.2019.00087>.

Xie, D.-Y., Sharma, S. B., Paiva, N. L., Ferreira, D., and Dixon, R. A. (2000). Role of Anthocyanidin Reductase, Encoded by Banyuls in Plant Flavonoid Biosynthesis. 18. V. Citovsky, Philos. Trans. R. Soc. London Ser. B (Vol. 289). www.sciencemag.org/cgi/content/full/299/5605/392/.

Zhang, X., Hu, D. peng, Li, Y., Chen, Y., Abidallha, E. H. M. A., Dong, Z. di, Chen, D. hua, and Zhang, L. (2017). Developmental and Hormonal Regulation of Fiber Quality in Two Natural-Colored Cotton Cultivars. Journal of Integrative Agriculture, 16(8), 1720–1729. [https://doi.org/10.1016/S2095-3119\(16\)61504-6](https://doi.org/10.1016/S2095-3119(16)61504-6).

Zhu, Y., Wang, H., Peng, Q., Tang, Y., Xia, G., Wu, J., and Xie, D. Y. (2015). Functional characterization of an anthocyanidin reductase gene from the fibers of upland cotton (*Gossypium hirsutum*). Planta, 241(5), 1075–1089. <https://doi.org/10.1007/s00425-014-2238-4>.

Zinn, R. A., and Owens, F. N. (1993). Ruminant Escape Protein for Lightweight Feedlot Calves. Journal Of Animal Science, 71(7), 1677–1687. <https://doi.org/10.2527/1993.7171677x>.