

DAFTAR PUSTAKA

- Adams, A. S., Currie, C. R., Cardoza, Y., Klepzig, K. D., & Raffa, K. F. (2009). Effects of symbiotic bacteria and tree chemistry on the growth and reproduction of bark beetle fungal symbionts. *Canadian Journal of Forest Research*, 39(6), 1133–1147. <https://doi.org/10.1139/X09-034>
- Arroyo-Manzanares, N., Campillo, N., López-García, I., & Viñas, P. (200 C.E.). *Determination of Aflatoxins by Liquid Chromatography Coupled to High-Resolution Mass Spectrometry*. www.intechopen.com
- Aryal, B., Adhikari, B., Aryal, N., Bhattarai, B. R., Khadayat, K., & Parajuli, N. (2021). LC-HRMS Profiling and Antidiabetic, Antioxidant, and Antibacterial Activities of *Acacia catechu* (L.f.) Willd. *BioMed Research International*, 2021. <https://doi.org/10.1155/2021/7588711>
- Asaf, S., Numan, M., Khan, A. L., & Al-Harrasi, A. (2020). Sphingomonas: from diversity and genomics to functional role in environmental remediation and plant growth. In *Critical Reviews in Biotechnology* (Vol. 40, Issue 2, pp. 138–152). Taylor and Francis Ltd. <https://doi.org/10.1080/07388551.2019.1709793>
- Bastías, D. A., Johnson, L. J., & Card, S. D. (2020). Symbiotic bacteria of plant-associated fungi: friends or foes? In *Current Opinion in Plant Biology* (Vol. 56, pp. 1–8). Elsevier Ltd. <https://doi.org/10.1016/j.pbi.2019.10.010>
- Behnouch, B., Sheikazadi, A., Bazmi, E., Fattahi, A., Sheikazadi, E., & Saberi Anary, S. H. (2015). Comparison of UHPLC and HPLC in benzodiazepines analysis of postmortem samples. *Medicine (United States)*, 94(14). <https://doi.org/10.1097/MD.0000000000000640>
- Beisser, D., Bock, C., Hahn, M. W., Vos, M., Sures, B., Rahmann, S., & Boenigk, J. (2019). Interaction-specific changes in the transcriptome of polynucleobacter asymbioticus caused by varying protistan communities. *Frontiers in Microbiology*, 10(JULY). <https://doi.org/10.3389/fmicb.2019.01498>
- Benkerroum, N. (2020). Chronic and acute toxicities of aflatoxins: Mechanisms of action. In *International Journal of Environmental Research and Public Health* (Vol. 17, Issue 2). MDPI AG. <https://doi.org/10.3390/ijerph17020423>
- Boersma, F. G. H., Warmink, J. A., Andreote, F. A., & van Elsas, J. D. (2009). Selection of Sphingomonadaceae at the base of *Laccaria proxima* and *Russula*

- exalbicans fruiting bodies. *Applied and Environmental Microbiology*, 75(7), 1979–1989. <https://doi.org/10.1128/AEM.02489-08>
- Brauer, V. S., Pessoni, A. M., Bitencourt, T. A., Paula, R. G. D., Rocha, L. D. O., Goldman, H. G., and Almeida, D., 2020. *Extracellular Vesicles from *Aspergillus flavus* Induce MIPolarization In Vitro*. *American Society For Microbiology*. 5(3): 1-9
<https://journals.asm.org/doi/epub/10.1128/mSphere.00190-20>
- Cabrera-Meraz, J., Maldonado, L., Bianchini, A., & Espinal, R. (2021). Incidence of aflatoxins and fumonisins in grain, masa and corn tortillas in four municipalities in the department of Lempira, Honduras. *Heliyon*, 7(12).
<https://doi.org/10.1016/j.heliyon.2021.e08506>
- Campos-Avelar, I., de la Noue, A. C., Durand, N., Cazals, G., Martinez, V., Strub, C., Fontana, A., & Schorr-Galindo, S. (2021). *Aspergillus flavus* growth inhibition and aflatoxin b1 decontamination by streptomyces isolates and their metabolites. *Toxins*, 13(5). <https://doi.org/10.3390/toxins13050340>
- Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K., Fierer, N., Pêa, A. G., Goodrich, J. K., Gordon, J. I., Huttley, G. A., Kelley, S. T., Knights, D., Koenig, J. E., Ley, R. E., Lozupone, C. A., McDonald, D., Muegge, B. D., Pirrung, M., ... Knight, R. (2010). QIIME allows analysis of high-throughput community sequencing data. In *Nature Methods* (Vol. 7, Issue 5, pp. 335–336). <https://doi.org/10.1038/nmeth.f.303>
- Chawla, G., & Ranjan, C. (2016). Principle, Instrumentation, and Applications of UPLC: A Novel Technique of Liquid Chromatography. *Open Chemistry Journal*, 3(1), 1–16. <https://doi.org/10.2174/1874842201603010001>
- Chen, Y. L., Lee, C. C., Lin, Y. L., Yin, K. M., Ho, C. L., & Liu, T. (2015). Obtaining long 16S rDNA sequences using multiple primers and its application on dioxin-containing samples. *BMC Bioinformatics*, 16(18).
<https://doi.org/10.1186/1471-2105-16-S18-S13>
- Debegnach, F., Brera, C., Mazzilli, G., Sonogo, E., Buiarelli, F., Ferri, F., Rossi, P. G., Collini, G., & de Santis, B. (2020). Optimization and validation of a LC-HRMS method for aflatoxins determination in urine samples. *Mycotoxin Research*, 36(2), 257–266. <https://doi.org/10.1007/s12550-020-00389-6>
- Deveau, A., Bonito, G., Uehling, J., Paoletti, M., Becker, M., Bindschedler, S., Hacquard, S., Hervé, V., Labbé, J., Lastovetsky, O. A., Mieszkin, S., Millet, L.

- J., Vajna, B., Junier, P., Bonfante, P., Krom, B. P., Olsson, S., van Elsas, J. D., & Wick, L. Y. (2018). Bacterial-fungal interactions: Ecology, mechanisms and challenges. In *FEMS Microbiology Reviews* (Vol. 42, Issue 3, pp. 335–352). Oxford University Press. <https://doi.org/10.1093/femsre/fuy008>
- Dewi, M. K., Ratnasari, E., Trimulyono, G., Biologi, J., Matematika, F., Pengetahuan, I., Universitas, A., & Surabaya, N. (n.d.). *Aktivitas Antibiotik Ekstrak Daun Majapahit (*Crescentia cujete*) terhadap Pertumbuhan Bakteri *Ralstonia solanacearum* Penyebab Penyakit Layu Antibacterial Activity of Majapahit (*Crescentia cujete*) Leaves Extract on *Ralstonia solanacearum**. <http://ejournal.unesa.ac.id/index.php/lenterabio>
- Edgar, R. C. (2017). Accuracy of microbial community diversity estimated by closed- and open-reference OTUs. *PeerJ*, 2017(10). <https://doi.org/10.7717/peerj.3889>
- Elise, M., Stéphanie, L., Leslie, A., Pascal, A., Jean-Marc, C., Vincent, F., Benoit, B., Eric, D., Jacques, G., Jean-Winoc, D., Anne, B., & Françoise, B. (2016). Characteristics of *aspergillus fumigatus* in association with *stenotrophomonas maltophilia* in an in vitro model of mixed biofilm. *PLoS ONE*, 11(11). <https://doi.org/10.1371/journal.pone.0166325>
- Fernández-Juri, M. G., Muzzolón, J. A., Dalcerro, A. M., & Magnoli, C. E. (2011). Effect of acid lactic bacteria isolated from faeces of healthy dogs on growth parameters and aflatoxin B1 production by *Aspergillus* species in vitro. *Mycotoxin Research*, 27(4), 273–280. <https://doi.org/10.1007/s12550-011-0104-9>
- Fong, A., & Shu, G. (2021). *Measurement of aflatoxin in maize/corn meal and other agricultural products*. 32. <https://doi.org/10.1117/12.2588282>
- Frey-Klett, P., Burlinson, P., Deveau, A., Barret, M., Tarkka, M., & Sarniguet, A. (2011). Bacterial-Fungal Interactions: Hyphens between Agricultural, Clinical, Environmental, and Food Microbiologists. *Microbiology and Molecular Biology Reviews*, 75(4), 583–609. <https://doi.org/10.1128/membr.00020-11>
- Frisvad, J. C., Skouboe, P., & Samson, R. A. (2005). Taxonomic comparison of three different groups of aflatoxin producers and a new efficient producer of aflatoxin B1, sterigmatocystin and 3-O-methylsterigmatocystin, *Aspergillus rambellii* sp. nov. *Systematic and Applied Microbiology*, 28(5), 442–453. <https://doi.org/10.1016/j.syapm.2005.02.012>

- Gerbaldo, G. A., Barberis, C., Pascual, L., Dalcero, A., & Barberis, L. (2012). Antifungal activity of two *Lactobacillus* strains with potential probiotic properties. *FEMS Microbiology Letters*, 332(1), 27–33. <https://doi.org/10.1111/j.1574-6968.2012.02570.x>
- Giang, L. T., & Thien, T. L. T. (2020). Determination of aflatoxin B1, B2, G1, G2 in cashew nut by UHPLC-HRMS. *Vietnam Journal of Chemistry*, 58(4), 540–547. <https://doi.org/10.1002/vjch.202000027>
- Gong, A. D., Lei, Y. Y., He, W. J., Liao, Y. C., Ma, L., Zhang, T. T., & Zhang, J. B. (2022). The Inhibitory Effect of *Pseudomonas stutzeri* YM6 on *Aspergillus flavus* Growth and Aflatoxins Production by the Production of Volatile Dimethyl Trisulfide. *Toxins*, 14(11). <https://doi.org/10.3390/toxins14110788>
- Gong, A. D., Wu, N. N., Kong, X. W., Zhang, Y. M., Hu, M. J., Gong, S. J., Dong, F. Y., Wang, J. H., Zhao, Z. Y., & Liao, Y. C. (2019). Inhibitory effect of volatiles emitted from *alcaligenes faecalis* N1-4 on *aspergillus flavus* and aflatoxins in storage. *Frontiers in Microbiology*, 10(JUN). <https://doi.org/10.3389/fmicb.2019.01419>
- Granillo, A. R., Canales, M. G. M., Espíndola, M. E. S., Rivera, M. A. M., de Lucio, V. M. B., & Tovar, A. V. R. (2015). Antibiosis interaction of *Staphylococcus aureus* on *Aspergillus fumigatus* assessed in vitro by mixed biofilm formation. *BMC Microbiology*, 15(1). <https://doi.org/10.1186/s12866-015-0363-2>
- Hamad, G. M., Zahran, E., & Hafez, E. E. (2017). The efficacy of bacterial and yeasts strains and their combination to bind aflatoxin B1 and B2 in artificially contaminated infants food. *Journal of Food Safety*, 37(4). <https://doi.org/10.1111/jfs.12365>
- He, Y., Caporaso, J. G., Jiang, X.-T., Sheng, H.-F., Huse, S. M., Rideout, J. R., Edgar, R. C., Kopylova, E., Walters, W. A., Knight, R., & Zhou, H.-W. (2015). Stability of operational taxonomic units: an important but neglected property for analyzing microbial diversity. *Microbiome*, 3(1). <https://doi.org/10.1186/s40168-015-0081-x>
- Hedayati, M. T., Pasqualotto, A. C., Warn, P. A., Bowyer, P., & Denning, D. W. (2007). *Aspergillus flavus*: Human pathogen, allergen and mycotoxin producer. In *Microbiology* (Vol. 153, Issue 6, pp. 1677–1692). <https://doi.org/10.1099/mic.0.2007/007641-0>

- Herlemann, D. P. R., Labrenz, M., Jürgens, K., Bertilsson, S., Waniek, J. J., & Andersson, A. F. (2011). Transitions in bacterial communities along the 2000 km salinity gradient of the Baltic Sea. *ISME Journal*, 5(10), 1571–1579. <https://doi.org/10.1038/ismej.2011.41>
- Herlinda, S., Darma Utama, M., Pujiastuti, Y., & Suwandi, D. (n.d.). *Kerapatan Dan Viabilitas Spora Beauveria Bassiana (Bals.) Akibat Subkultur Dan Pengayaan Media, Serta Virulensinya Terhadap Larva Plutella Xylostella (Linn.)*.
- Hogan, D. A., & Kolter, R. (2002). Pseudomonas-Candida Interactions: An Ecological Role for Virulence Factors. In *Source: Science, New Series* (Vol. 296, Issue 5576).
- Homa, M., Sándor, A., Tóth, E., Szebenyi, C., Nagy, G., Vágvölgyi, C., & Papp, T. (2019). In vitro interactions of Pseudomonas aeruginosa with scedosporium species frequently associated with cystic fibrosis. *Frontiers in Microbiology*, 10(MAR). <https://doi.org/10.3389/fmicb.2019.00441>
- Hyde, K. D., Al-Hatmi, A. M. S., Andersen, B., Boekhout, T., Buzina, W., Dawson, T. L. J. R., Eastwood, D. C., Jones, E. B. G., De Hoog, S., Kang, Y., Longcore, E. J., McKenzie, E. H. C., Meis, J. F., Pinson-Gadais, L., Rathnayaka, A.R., Richard-Forget, F., Stadler, M., Theelen, B., Thongbai, B., and Tsui, C. K. M. (2018). *The world's ten most feared fungi*. *Fungal Divers* 9: 161–194 <https://link.springer.com/article/10.1007/s13225-018-0413-9>
- Idiyatov, I. I., Eroshin, A. I., Yusupov, S. A., Tremasova, A. M., & Biryulya, V. v. (2022). Endophytic bacteria antagonists of the micromycete *Aspergillus flavus*: The prospect of improving the quality of food raw materials and food products. *IOP Conference Series: Earth and Environmental Science*, 949(1). <https://doi.org/10.1088/1755-1315/949/1/012072>
- Jia, K., Yan, L., Jia, Y., Xu, S., Yan, Z., & Wang, S. (2021). Afln is involved in the biosynthesis of aflatoksin and conidiation in *Aspergillus flavus*. *Toxins*, 13(11). <https://doi.org/10.3390/toxins13110831>
- Katzung, Betram G. (2010). *Farmakologi Dasar dan Klinik*. Edisi 10. EGC, Jakarta.
- Kasno, A. (2004). Balai Penelitian Tanaman Kacang-kacangan dan Umbi-umbian. In *Jurnal Litbang Pertanian* 23 (3) : 1-7 <http://203.190.36.42/publikasi/p3233041.pdf>

- Khattak, S. U., Lutfullah, G., Iqbal, Z., Ahmad, J., Rehman, I. U., Shi, Y., & Ikram, S. (2021). *Aspergillus flavus* originated pure compound as a potential antibacterial. *BMC Microbiology*, 21(1). <https://doi.org/10.1186/s12866-02102371-3>
- Klich, M. A. (2007). *Aspergillus flavus: the major producer of aflatoxin*, *Molecular Plant Pathology*, 8(6): 713–722
<https://bsppjournals.onlinelibrary.wiley.com/doi/epdf/10.1111/j.13643703.2007.00436.x>
- Klindworth, A., Pruesse, E., Schweer, T., Peplies, J., Quast, C., Horn, M., & Glöckner, F. O. (2013). Evaluation of general 16S ribosomal RNA gene PCR primers for classical and next-generation sequencing-based diversity studies. *Nucleic Acids Research*, 41(1). <https://doi.org/10.1093/nar/gks808>
- Kolawole, O., Meneely, J., Petchkongkaew, A., & Elliott, C. (2021). A review of mycotoxin biosynthetic pathways: associated genes and their expressions under the influence of climatic factors. In *Fungal Biology Reviews* (Vol. 37, pp. 8–26). Elsevier Ltd. <https://doi.org/10.1016/j.fbr.2021.04.003>
- Krishnan, S., Manavathu, E. K., & Chandrasekar, P. H. (2009). *Aspergillus flavus*: An emerging non-fumigatus *Aspergillus* species of significance. In *Mycoses* (Vol. 52, Issue 3, pp. 206–222). <https://doi.org/10.1111/j.1439-0507.2008.01642.x>
- Kumar, V., Bahuguna, A., Ramalingam, S., Dhakal, G., Shim, J. J., & Kim, M. (2022). Recent technological advances in mechanism, toxicity, and food perspectives of enzyme-mediated aflatoxin degradation. In *Critical Reviews in Food Science and Nutrition* (Vol. 62, Issue 20, pp. 5395–5412). Taylor and Francis Ltd. <https://doi.org/10.1080/10408398.2021.2010647>
- Li, Y., Liu, Y. H., Yang, Z. B., Wan, X. L., & Chi, F. (2012). The efficacy of clay enterosorbent to ameliorate the toxicity of aflatoxin B1 from contaminated corn (*zea mays*) on hematology, serum biochemistry, and oxidative stress in ducklings. *Journal of Applied Poultry Research*, 21(4), 806–815. <https://doi.org/10.3382/japr.2012-00538>
- Li, L., Ma, M., Huang, R., Qu, Q., Li, G., Zhou, J., Zhang, K., Lu, K., Niu, X., & Luo, J. (2012). Induction of chlamydospore formation in fusarium by cyclic lipopeptide antibiotics from bacillus subtilis C2. *Journal of Chemical Ecology*, 38(8), 966–974. <https://doi.org/10.1007/s10886-012-0171-1>

- Li, S., Xu, X., Zhao, T., Ma, J., Zhao, L., Song, Q., & Sun, W. (2022). Screening of *Bacillus velezensis* E2 and the Inhibitory Effect of Its Antifungal Substances on *Aspergillus flavus*. *Foods*, *11*(2). <https://doi.org/10.3390/foods11020140>
- Lladó Fernández, S., Větrovský, T., & Baldrian, P. (2019). The concept of operational taxonomic units revisited: genomes of bacteria that are regarded as closely related are often highly dissimilar. *Folia Microbiologica*, *64*(1), 19–23. <https://doi.org/10.1007/s12223-018-0627-y>
- Leveau, J. H. J., & Preston, G. M. (2008). Bacterial mycophagy: Definition and diagnosis of a unique bacterial-fungal interaction. In *New Phytologist* (Vol. 177, Issue 4, pp. 859–876). <https://doi.org/10.1111/j.1469-8137.2007.02325.x>
- Majumdar, R., Kandel, S. L., Cary, J. W., & Rajasekaran, K. (2021). Changes in bacterial endophyte community following *aspergillus flavus* infection in resistant and susceptible maize kernels. *International Journal of Molecular Sciences*, *22*(7). <https://doi.org/10.3390/ijms22073747>
- Martindah, E., & Bahri, S. (2017). Mycotoxin Contamination in the Food Chain. *Indonesian Bulletin of Animal and Veterinary Sciences*, *26*(3), 115. <https://doi.org/10.14334/wartazoa.v26i3.1393>
- Massomo, S. M. S. (2020). *Aspergillus flavus* and aflatoxin contamination in the maize value chain and what needs to be done in Tanzania. In *Scientific African* (Vol. 10). Elsevier B.V. <https://doi.org/10.1016/j.sciaf.2020.e00606>
- Mitchell, N. J., Bowers, E., Hurburgh, C., & Wu, F. (2016). Potential economic losses to the US corn industry from aflatoxin contamination. *Food Additives and Contaminants - Part A Chemistry, Analysis, Control, Exposure and Risk Assessment*, *33*(3), 540–550. <https://doi.org/10.1080/19440049.2016.1138545>
- Muthukrishnan, S., Palanisamy, S., Subramanian, S., Selvaraj, S., Mari, K. R., & Kupplingam, R. (2016). Phytochemical Profile of *Erythrina variegata* by Using High-Performance Liquid Chromatography and Gas Chromatography-Mass Spectroscopy Analyses. *JAMS Journal of Acupuncture and Meridian Studies*, *9*(4), 207–212. <https://doi.org/10.1016/j.jams.2016.06.001>
- Nakatsu, C. H., Byappanahalli, M. N., & Nevers, M. B. (2019). Bacterial community 16S rRNA gene sequencing characterizes riverine microbial impact on Lake Michigan. *Frontiers in Microbiology*, *10*(MAY). <https://doi.org/10.3389/fmicb.2019.00996>

- Namjoo, M., Salamat, F., Rajabli, N., Haji-Hoseeini, R., Niknejad, F., Kohsar, F., & Joshaghani, H. (2016). Quantitative Determination of Aflatoxin by High Performance Liquid Chromatography in Wheat Silos in Golestan Province, North of Iran. In *Iran J Public Health* (Vol. 45, Issue 7). <http://ijph.tums.ac.ir>
- Nasir, N., Sayeed, M. A., & Jamil, B. (2019). *Ralstonia pickettii* Bacteremia: An Emerging Infection in a Tertiary Care Hospital Setting. *Cureus*. <https://doi.org/10.7759/cureus.5084>
- Negi, J. S., Singh, P., Pant, G. J. N., & Rawat, M. S. M. (2011). High-performance liquid chromatography analysis of plant saponins: An update 2005-2010. In *Pharmacognosy Reviews* (Vol. 5, Issue 10, pp. 155–158). <https://doi.org/10.4103/0973-7847.91109>
- Nováková, L., Solichová, D., & Solich, P. (2006). Advantages of ultra performance liquid chromatography over high-performance liquid chromatography: Comparison of different analytical approaches during analysis of diclofenac gel. *Journal of Separation Science*, 29(16), 2433–2443. <https://doi.org/10.1002/jssc.200600147>
- O'Brien, J., & Wright, G. D. (2011). An ecological perspective of microbial secondary metabolism. In *Current Opinion in Biotechnology* (Vol. 22, Issue 4, pp. 552–558). <https://doi.org/10.1016/j.copbio.2011.03.010>
- Pasikanti, K. K., Ho, P. C., & Chan, E. C. Y. (2008). Gas chromatography/mass spectrometry in metabolic profiling of biological fluids. In *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences* (Vol. 871, Issue 2, pp. 202–211). <https://doi.org/10.1016/j.jchromb.2008.04.033>
- Partida-Martinez, L. P., Monajembashi, S., Greulich, K. O., & Hertweck, C. (2007). Endosymbiont-Dependent Host Reproduction Maintains Bacterial-Fungal Mutualism. *Current Biology*, 17(9), 773–777. <https://doi.org/10.1016/j.cub.2007.03.039>
- Peach, K. C., Bray, W. M., Winslow, D., Linington, P. F., & Linington, R. G. (2013). Mechanism of action-based classification of antibiotics using high-content bacterial image analysis. *Molecular BioSystems*, 9(7), 1837–1848. <https://doi.org/10.1039/c3mb70027e>

- Pelczar MJ & Chan ECS, 1988. *Dasar-Dasar Mikrobiologi 2*. Jakarta: Penerbit Universitas Indonesia
- Peleg, A. Y., Hogan, D. A., & Mylonakis, E. (2010). Medically important bacterial-fungal interactions. In *Nature Reviews Microbiology* (Vol. 8, Issue 5, pp. 340–349). <https://doi.org/10.1038/nrmicro2313>
- Peleg, A. Y., Tampakakis, E., Burgwyn Fuchs, B., Eliopoulos, G. M., Moellering, R. C., & Mylonakis, E. (2008). *Prokaryote-eukaryote interactions identified by using *Caenorhabditis elegans**. www.pnas.org/cgi/content/full/
- Peterson, S. W., Ito, Y., Horn, B. W., & Goto, T. (2001). *Aspergillus bombycis*, a new aflatoxigenic species and genetic variation in its sibling species, *A. nomius*. *Mycologia*, 93(4), 689–703. <https://doi.org/10.1080/00275514.2001.12063200>
- Pierce, E. C., Morin, M., Little, J. C., Liu, R. B., Tannous, J., Keller, N. P., Pogliano, K., Wolfe, B. E., Sanchez, L. M., & Dutton, R. J. (2021). Bacterial–fungal interactions revealed by genome-wide analysis of bacterial mutant fitness. *Nature Microbiology*, 6(1), 87–102. <https://doi.org/10.1038/s41564-02000800-z>
- Pinto-Tomás, A. A., Anderson, M. A., Suen, G., Stevenson, D. M., Chu, F. S. T., Wallace Cleland, W., Weimer, P. J., & Currie, C. R. (2009). Symbiotic nitrogen fixation in the fungus gardens of leaf-cutter ants. *Science*, 326(5956), 1120–1123. <https://doi.org/10.1126/science.1173036>
- Poretzky, R., Rodriguez-R, L. M., Luo, C., Tsementzi, D., & Konstantinidis, K. T. (2014). Strengths and limitations of 16S rRNA gene amplicon sequencing in revealing temporal microbial community dynamics. *PLoS ONE*, 9(4). <https://doi.org/10.1371/journal.pone.0093827>
- Purnomo, A. S., Sariwati, A., & Kamei, I. (2020). Synergistic interaction of a consortium of the brown-rot fungus *Fomitopsis pinicola* and the bacterium *Ralstonia pickettii* for DDT biodegradation. *Heliyon*, 6(6). <https://doi.org/10.1016/j.heliyon.2020.e04027>
- Rangel-Muñoz, E. J., Valdivia-Flores, A. G., Moreno-Rico, O., Hernández-Delgado, S., Cruz-Vázquez, C., de-Luna-López, M. C., Quezada-Tristán, T., Ortiz-Martínez, R., & Máyek-Pérez, N. (2020). Caracterización de *Aspergillus flavus* y cuantificación de aflatoxinas en pienso y leche cruda de vacas en

Aguascalientes, México. *Revista Mexicana De Ciencias Pecuarias*, 11(2), 435–454. <https://doi.org/10.22319/RMCP.V11I2.5686>

Ranjard, L., Brothier, E., & Nazaret, S. (2000). Sequencing bands of ribosomal intergenic spacer analysis fingerprints for characterization and microscale distribution of soil bacterium populations responding to mercury spiking. *Applied and Environmental Microbiology*, 66(12), 5334–5339. <https://doi.org/10.1128/AEM.66.12.5334-5339.2000>

Reuhs, B. L., & Rounds, M. A. (2010). *High-Performance Liquid Chromatography* (pp. 499–512). https://doi.org/10.1007/978-1-4419-1478-1_28

Robinson, A. J., House, G. L., Morales, D. P., Kelliher, J. M., Gallegos-Graves, L. V., LeBrun, E. S., Davenport, K. W., Palmieri, F., Lohberger, A., Bregnard, D., Estoppey, A., Buffi, M., Paul, C., Junier, T., Hervé, V., Cailleau, G., Lupini, S., Nguyen, H. N., Zheng, A. O., ... Chain, P. S. G. (2021). Widespread bacterial diversity within the bacteriome of fungi. *Communications Biology*, 4(1). <https://doi.org/10.1038/s42003-021-02693-y>

Romero-Sánchez, I., Ramírez-García, L., Gracia-Lor, E., & Madrid-Albarrán, Y. (2022). Simultaneous determination of aflatoxins B1, B2, G1 and G2 in commercial rices using immunoaffinity column clean-up and HPLC-MS/MS. *Food Chemistry*, 395. <https://doi.org/10.1016/j.foodchem.2022.133611>

Samson, R. A., Visagie, C. M., Houbraeken, J., Hong, S. B., Hubka, V., Klaassen, C. H. W., Perrone, G., Seifert, K. A., Susca, A., Tanney, J. B., Varga, J., Kocsubé, S., Szigeti, G., Yaguchi, T., & Frisvad, J. C. (2014). Phylogeny, identification and nomenclature of the genus *Aspergillus*. *Studies in Mycology*, 78(1), 141–173. <https://doi.org/10.1016/j.simyco.2014.07.004>

Sayed, E. T., Desouky, E. T., & el Aziz, A. A. (n.d.). *Investigation of fungus associated within co-occurrence of aflatoxins and ochratoxin a in cereals from Egypt*. <https://doi.org/10.15406/mojt.2019.05.00161>

Schulz-Bohm, K., Tyc, O., de Boer, W., Peereboom, N., Debets, F., Zaagman, N., Janssens, T. K. S., & Garbeva, P. (2017). Fungus-associated bacteriome in charge of their host behavior. *Fungal Genetics and Biology*, 102, 38–48. <https://doi.org/10.1016/j.fgb.2016.07.011>

Siddiqui, N. A. (n.d.). *The Effect of *Alcaligenes faecalis* on Inhibition of *Candida albicans* Biofilm and Planktonic Growth*. <https://dc.etsu.edu/honors/575>

- Spraker, J. E., Sanchez, L. M., Lowe, T. M., Dorrestein, P. C., & Keller, N. P. (2016). *Ralstonia solanacearum* lipopeptide induces chlamydospore development in fungi and facilitates bacterial entry into fungal tissues. *ISME Journal*, *10*(9), 2317–2330. <https://doi.org/10.1038/ismej.2016.32>
- Steffan, B. N., Venkatesh, N., & Keller, N. P. (2020). Let's get physical: Bacterial-fungal interactions and their consequences in agriculture and health. In *Journal of Fungi* (Vol. 6, Issue 4, pp. 1–18). MDPI AG. <https://doi.org/10.3390/jof6040243>
- Sutherland, R., & Rolinson, G. N. (1964). Activity Of Ampicillin In Vitro Compared With Other Antibiotics. *Journal of Clinical Pathology*, *17*, 461–465. <https://doi.org/10.1136/jcp.17.4.461>
- Tampakakis, E., Peleg, A. Y., & Mylonakis, E. (2009). Interaction of candida albicans with an intestinal pathogen, salmonella enterica serovar typhimurium. *Eukaryotic Cell*, *8*(5), 732–737. <https://doi.org/10.1128/EC.00016-09>
- Thijs, S., de Beeck, M. O., Beckers, B., Truyens, S., Stevens, V., van Hamme, J. D., Weyens, N., & Vangronsveld, J. (2017). Comparative evaluation of four bacteria-specific primer pairs for 16S rRNA gene surveys. *Frontiers in Microbiology*, *8*(MAR). <https://doi.org/10.3389/fmicb.2017.00494>
- Tropika, A. (2020). *Ability Test Of Trichoderma Spp. Isolate As Antagonist Of Ganoderma Boninense And Plant Growth Promoting Fungi (Pgpf)* (Vol. 8, Issue 2).
- Uka, V., Cary, J. W., Lebar, M. D., Puel, O., de Saeger, S., & Diana Di Mavungu, J. (2020). Chemical repertoire and biosynthetic machinery of the *Aspergillus flavus* secondary metabolome: A review. *Comprehensive Reviews in Food Science and Food Safety*, *19*(6), 2797–2842. <https://doi.org/10.1111/15414337.12638>
- Wang, T., & Zhang, Q. (2021). Production of monoclonal antibodies against AFLM (Ver-1), a middle key protein involved in aflatoksin biosynthesis. *Oil Crop Science*, *6*(4), 201–205. <https://doi.org/10.1016/j.ocsci.2021.10.003>
- Yan, Q., Zhou, J., Li, H., Zhi, Q., Zhou, X., He, Z., 2015. Coexistence of and Interaction Relationships between an Aflatoksin-producing Fungus and a Bacterium. *Fungal Biology. Hal* : 1-36

- Yang, M., Zhu, Z., Zhuang, Z., Bai, Y., Wang, S., & Ge, F. (2021). Proteogenomic characterization of the pathogenic fungus *aspergillus flavus* reveals novel genes involved in aflatoksin production. *Molecular and Cellular Proteomics*, 20. <https://doi.org/10.1074/MCP.RA120.002144>
- Yergeau, E., Bell, T. H., Champagne, J., Maynard, C., Tardif, S., Tremblay, J., & Greer, C. W. (2015). Transplanting soil microbiomes leads to lasting effects on willow growth, but not on the rhizosphere microbiome. *Frontiers in Microbiology*, 6(DEC). <https://doi.org/10.3389/fmicb.2015.01436>
- Yu, J., Cleveland, T. E., Nierman, W. C., & Bennett, J. W. (2005). *Aspergillus flavus* genomics: Gateway to human and animal health, food safety, and crop resistance to diseases. In *Revista Iberoamericana de Micologia* (Vol. 22, Issue 4, pp. 194–202). Asociacion Espanola de Micologia. [https://doi.org/10.1016/S1130-1406\(05\)70043-7](https://doi.org/10.1016/S1130-1406(05)70043-7)
- Zhang, L., Kang, M., Huang, Y., & Yang, L. (2016). Fungal communities from the calcareous deep-sea sediments in the Southwest India Ridge revealed by Illumina sequencing technology. *World Journal of Microbiology and Biotechnology*, 32(5). <https://doi.org/10.1007/s11274-016-2030-7>
- Zhang, J., Wang, Y., Du, Z., Lin, D., Huo, L., Qin, L., Wang, W., Qiang, L., Yao, Y., & An, Y. (2021). Screening, identification and antagonistic effect of antagonistic bacteria JTFM1001 against aflatoksin contamination in corn. *Oil Crop Science*, 6(1), 1–<https://doi.org/10.1016/j.ocsci.2021.01.003>