



## REFERENCES

- Alastruey-Izquierdo, A., Melhem, M. S. Bonfietti, L. X., Rodriguez-Tudela, J.L. (2015). Susceptibility test for fungi: clinical and laboratorial correlations in medical mycology. *Revista do Instituto de Medicina Tropical de São Paulo*, 57 Suppl 19 (Suppl 19), 57–64. <https://doi.org/10.1590/S0036-46652015000700011>
- Alen, Y., Amelia, R., Djamaan, A. (2018). TLC profile and activity test of secondary metabolites Aspergillus flavus "in-habiting" queen termite's nest *Macrotermes gilvus* on enriched media. *Indonesian Journal of Pharmaceutical Science and Technology*, 5(1): 31-42. <http://jurnal.unpad.ac.id/ijpst/article/view/14644>
- Ali, S., Khan, M.R., Batool, R., Maryam, S., Majid, M. (2019). Wound healing potential of oil extracted from *Parrotiopsis jacquemontiana* (Decne) Rehder. *J. Ethnopharmacol*, 236: 354-365. <https://doi.org/10.1016/j.jep.2019.03.018>
- Al-Shubaib, Mohammed Baqir & Albakri, Ali & Hashim, Hayder & Alwan, Sabah & Almandil, Noor & Selvaraj, Prathas & Jermy, Rabindran & Abdulazeez, Sayed & Borgio, J Francis. (2020). High Prevalence of Aflatoxin B 1 in Aspergillus flavus Infecting Stored Rice Grains. *J. Agr. Sci. Tech*, 22. 737-745.
- Anonim (2021). Termite pictures-*Macrotermes givus* Termite Web. <https://www.termiteweb.com/termite-pictures-macrotermes-gilvus/>
- Arifin, Z., Dahlan, Z., Sabaruddin, Irsan, C., Hartono, Y. (2014). Characteristics, morphometry and spatial distribution of populations of subterranean termites *Macrotermes gilvus*. Hagen (Isoptera: Termitidae) in the rubber plantation land habitat which was managed without pesticides and chemical fertilizers. *International Journal of Science and Research*, 3(4): 102-106
- Arinana, A.R., Nandika, D., Rauf, A., Harahap I.S., Sumertajaya, I.M., Bahtiar, E.T. (2016). Termite diversity in urban landscape, South Jakarta, Indonesia. *Insects*, 7: 1-18. <https://doi.org/10.3390/insects7020020>
- Azziz-Baumgartner, E., Lindblade, K., Gieseke, K., Rogers, H. S., Kieszak, S., Njapau, H., et al. (2005). Case-control study of an acute aflatoxicosis outbreak, Kenya, 2004. *Environ. Health Perspect.* 113, 1779–1783.
- Bandyopadhyay, R., and Cotty, P.L., (2013). Biological controls for aflatoxin reduction. Pages 44-45 in: Aflatoxins: Finding Solutions for Improved Food Safety. L. Unnevehr and D. Grace, eds. IFPRI, Washington DC
- Bbosa, G.S., Kitya, D., Lubega, A., Ogwali-Okeng, J., Anokbonggo, W.W., Kyegombe, D.B. (2013). Review of the biological and health effects of aflatoxins on body organs



and body systems. *Aflatoxins-Recent Advances and Future Prospects.*  
<https://doi.org/10.5772/51201>

Bernáldez, V., Córdoba, J. J., Magan, N., Peromingo, B., and Rodríguez, A. (2017). The influence of ecophysiological factors on growth, aflR gene expression and aflatoxin B1 production by a type strain of *Aspergillus flavus*. *LWT Food Sci. Technol*, 83, 283–291. <https://doi.org/10.1016/j.lwt.2017.05.030>

Boer SH.J., Boyd-Wilson K.S.H., Petley, M., Walter M . Influence of cold-storage temperatures on strawberry leak caused by *Rhizopus* spp. *N Z Plant Protect*, 2009;62:243–249.

Bui-Klimke, T.R., Wu, F. (2016). Ochratoxin A and human health risk: A review of the evidence. *Crit Rev Food Sci Nutr*, 55(13): 1860-1869.  
<https://doi.org/10.1080/10408398.2012.724480>

CLSI. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi; Approved Standard CLSI Document M38-A2*. Wayne, PA: Clinical and Laboratory Standards Institute; 2008

Cotty, P.J., Jaime-Garcia, R. (2007). Influences of climate on aflatoxin producing fungi and aflatoxin contamination. *Int J Food Microbiol*, 119(1-2):109-15.  
<https://doi.org/10.1016/j.ijfoodmicro.2007.07.060>.

Culliau, A. G. L. and Barcello, J. M. (2015). Fungal and mycotoxin contamination of coffee beans in Benguet province, Philippines, food additives & contaminants: Part A, 32(2), pp. 250–260. <https://doi.org/10.1080/19440049.2014.1001796>

Dar, Mohammad Shafi & Sreedar, Gadiputi & Shukla, Abhilasha & Gupta, Prashant & Rehan, AhmadDanish & George, Jiji. (2015). An in vitro study of antifungal drug susceptibility of Candida species isolated from human immunodeficiency virus seropositive and human immunodeficiency virus seronegative individuals in Lucknow population Uttar Pradesh. *Journal of Oral and Maxillofacial Pathology*. 19. 205. <https://doi.org/10.4103/0973-029X.164534>.

Dinev, T., Tzanova, M., Velichkova, ., Dermendzhieva, D., Beev, G. (2021). . Antifungal and Antioxidant Potential of Methanolic Extracts from *Acorus calamus* L., *Chlorella vulgaris* Beijerinck, *Lemna minuta* Kunth and *Scenedesmus dimorphus* (Turpin) Kützing. *Appl. Sci*, 11: 4745. <https://doi.org/10.3390/app11114745>

Enikova, Rositsa & Stoynovska, Mariyana & Karcheva, Milena. (2020). Mycotoxins in fruits and vegetables. *Journal of IMAB - Annual Proceeding (Scientific Papers)*. 26. <https://doi.org/10.5272/jimab.2020262.3139>.



- Ester, A., Mukarlina, dan Rahmawati. 2017. Aktivitas ekstrak metanol daun sembung rambat (*Mikania micrantha* Kunth) terhadap pertumbuhan *Phytophthora* sp. Im5 dari batang jeruk siam (*Citrus nobilis* var. *microcarpa*). *J. Protobiont.* 6(2): 63–67.
- Espinel-Ingroff, A., Fothergill, A., Peter, J., Rinaldi, M.G., Walsh, T.J. (2002). Testing conditions for determination of minimum fungicidal concentrations of new and established antifungal agent for *Aspergillus* spp.: NCCLS collaborative study. *Journal of Clinical Microbiology* 40(9): 3204-3208. <https://doi.org/10.1128/JCM.40.9.3204-3208.2002>
- Espinel-Ingroff, A., Fothergill, A., Peter, J., Rinaldi, M.G., Walsh, T.J. (2002). Quality control and reference guidelines for CLSI Broth Microdilution Method (M38-A Document) for susceptibility testing of anidulafungin against molds. *Journal of Clinical Microbiology* 45(7): 2180–2182. <https://doi.org/10.1128/JCM.00399-07>
- Espinel-Ingroff, Ana. (2003). In vitro antifungal activities of anidulafungin and micafungin, licensed agents and the investigational triazole posaconazole as determined by NCCLS methods for 12, 052 fungal isolates: review of the literature. *Rev Iberoam Micol*, 20(4):121-36. PMID: 15456349.
- EUCAST. 2008. EUCAST Definitive Document EDef 7.1: method for the determination of broth dilution MICs of antifungal agents for fermentative yeasts. *Clinical Microbiology and Infection* 14(4):398-405.
- FAO. 2019. *The State of Food and Agriculture 2019. Moving forward on food loss and waste reduction*. Rome: FAO
- FDA. (2003a). 21CFR170.30. *Elegibility for classification as generally recognised as safe (GRAS)*.
- FDA. (2003b). 21CFR 172.55. *Natamycin (pimaricin)*
- Feliziani, E., Romanazzi, G. 2016. Postharvest decay of strawberry fruit: Etiology, epidemiology, and disease management. *Journal of Berry Research*, vol. 6, no. 1, pp. 47-63
- Francisco, Castillo & Castillo, Francisco & Julio, Alberto & Gallegos M, Gabriel & I, Rodr & bal, No. (2015). In vitro antifungal activity of polyphenols-rich plant extracts against *Phytophthora cinnamomi* Rands. African Journal of Agricultural Research. <https://doi.org/10.4554-4560.10.5897/AJAR2013.8072>.
- Habib, M.R., Karim, M.R. (2009) Antimicrobial and cytotoxic activity of Di-(2-ethylhexyl) phthalate and anhydrosophoradiol-3-acetate isolated from *Calotropis gigantea* (Linn.) flower. *Mycobiology*. <https://doi.org/10.4489/MYCO.2009.37.1.031>



- Hollick GE, Washington JA., 2nd Comparison of direct and standardized disk diffusion susceptibility testing of urine cultures. *Antimicrob Agents Chemother.* 1976, 9:804–809. <https://doi.org/10.1128/AAC.9.5.804>.
- IARC (2002). *Monographs on the Evaluation of Carcinogenic Risks to Humans*, Vol. 82. Lyon: International Agency for Research on Cancer.
- Iqbal, S. Z., Jinap, S., Pirouz, A. A., and Faizal, A. R. A. (2015). Aflatoxin M1 in milk and dairy products, occurrence and recent challenges: a review. *Trends Food Sci. Technol.* 46, 110–119. <https://doi.org/10.1016/j.tifs.2015.08.005>
- Jaimez, J., Fente, C. A., Vazquez, B. I., Franco, C. M., Cepeda, A., Mahuzier, G. (2000). Application of the assay of aflatoxins by liquid chromatography with fluorescence detection in food analysis. *J. Chromatogr. A* 882, 1–10. [https://doi.org/10.1016/S0021-9673\(00\)00212-0](https://doi.org/10.1016/S0021-9673(00)00212-0)
- Hudzicki, Jan. (2009). *Kirby-Bauer Disk Diffusion Susceptibility Test Protocol: 8 Desember*. American Society for Microbiology. <https://asm.org/getattachment/2594ce26-bd44-47f6-8287-0657aa9185ad/Kirby-Bauer-Disk-Diffusion-Susceptibility-Test-Protocol-pdf.pdf.pdf>
- Jeff-Agboola, Y.A., Awe, L.B., (2016). Antifungal and phytochemical screening of some Nigerian medicinal plant extracts against toxigenic *Aspergillus flavus*, *Cogent Food & Agriculture*, 2:1, <https://doi.org/10.1080/23311932.2016.1210556>
- Johnson, E. M., Ojwang, J. O., Szekely, A., Wallace, T. L., & Warnock, D. W. (1998). Comparison of in vitro antifungal activities of free and liposome-encapsulated nystatin with those of four amphotericin B formulations. *Antimicrobial agents and chemotherapy*, 42(6), 1412–1416. <https://doi.org/10.1128/AAC.42.6.1412>
- Khan, R., Ghazali, F.R., Mahyudin, N.A., Samsudin, N.I.P. (2020). Morphological characterization and determination of aflatoxigenic and non-aflatoxigenic *Aspergillus flavus* isolated from sweet corn kernels and soil in Malaysia, *Agriculture*, 10:450; <https://doi.org/10.3390/agriculture10100450>
- Krishnamachari, K. A. V. R., Bhat, R. V., Nagarajan, V., and Tilak, T. B. G. (1975). Hepatitis due to aflatoxicosis-An outbreak in western India. *Lancet* 1, 1061–1063. [https://doi.org/10.1016/S0140-6736\(75\)91829-2](https://doi.org/10.1016/S0140-6736(75)91829-2)
- Konaté S., Roux, X.L., Verdier, B., Lepage, M. (2003). Effect of underground fungus-growing termites on carbon dioxide emissions at the point- and landscape-scales in an African savanna. *Journal of Functional Ecology*, 17(3): 305- 314. <https://doi.org/10.1046/j.1365-2435.2003.00727.x>



- Kumar, P., Mahato, D. K., Kamle, M., Mohanta, T. K., and Kang, S. G. (2017). Aflatoxins: a global concern for food safety, human health and their management. *Front. Microbiol.* 7:2170. <https://doi.org/10.3389/fmicb.2016.02170>
- Lightbown, J. W., Kogut, M., & Uemura, K. (1963). The international standard for nystatin. *Bulletin of the World Health Organization*, 29(1), 87–94.
- Lv, C., Jin, J., Wang, P., Dai, X., Liu, Y., Zheng, M., et al. (2019). Interaction of water activity and temperature on the growth, gene expression and aflatoxin production by *Aspergillus flavus* on paddy and polished rice. *Food Chem.* 293, 472–478. <https://doi.org/10.1016/j.foodchem.2019.05.009>
- Mahaney, W.C.; Zippin, J.; Milner, M.W.; Sanmugadas, K.; Kanckock, R.G.V.; Aufreiter, S. (1999). Chemistry, mineralogy and microbiology of termite mound soil eaten by chimpanzees of the Mahal mountains, Western Tanzania. *J. Trop. Eco.* 15, 565–588.
- Mahato, D.K., Lee K.E., Kamle, M., Devi, S., Dewangan, K.N., Kumar, P., Kang, S.G. (2019). Aflatoxins in food and feed: An overview on prevalence, detection, and controls strategies. *Front. Microbiol.* <https://doi.org/10.3389/fmicb.2019.02266>
- Meletiadis, J., Meis, J.F., Mouton, J.W., Verweij, P.E. (2001). Analysis of growth characteristics of filamentous fungi in different nutrient media. *J Clin Microbiol.* 39(2):478-84. <https://doi.org/10.1128/JCM.39.2.478-484.2001>. PMID: 11158093; PMCID: PMC87762.
- Methano. (2012). *Chemical structure of Nystatin*. [wikimedia SVG] Pub Chem. <https://commons.wikimedia.org/w/index.php?curid=21972878>
- Mattia, A., Sipes, G.I. WHO food additives series No. 46. *Inchem*. Retrieved January 7<sup>th</sup>, 2022, from [https://inchem.org/documents/jecfa/jecmono/v46je09.htm#\\_46091600](https://inchem.org/documents/jecfa/jecmono/v46je09.htm#_46091600)
- Manasseh, T.R., Zacchaeus, I.S., Gwatau, D.D., Falang, K.D., and Wuyep, K.A. (2020). Ethyl acetate root extract of *Terminalia glaucescens* protects *Drosophila melanogaster* against virulent *Aspergillus* Species. *Journal of Applied Sciences*, 20: 44-50.
- Miskiyah, M., Winarti, C., & Broto, W. (2016). Kontaminasi mikotoksin pada buah segar dan produk olahannya serta penanggulangannya. *Jurnal Penelitian dan Pengembangan Pertanian*, 29(3), 79-85.
- Mutegi, C. K., Ngugi, H. K., Hendriks, S. L., and Jones, R. B. (2009). Prevalence and factors associated with aflatoxin contamination of peanuts from Western Kenya. *Int. J. Food Microbiol.* 130, 27–34. <https://doi.org/10.1016/j.ijfoodmicro.2008.12.030>
- Lee, K.E., Wood, T.G. (1971). *Termite and Soil*; London Academic Press.



- Nakasugi, L.P., Bomfim, N.S., Romoli, J.C.Z., Neroli, S.B., Silva, M.V., Oliveira, G.H.R., Jr, M.M. (2021). Antifungal and anti aflatoxigenic activities of thymol and carvacrol against *Aspergillus flavus*. *Saud Pesq*, 14(1): 113-123. <https://doi.org/10.17765/2176-9206.2021c14n1.e7727>
- Nandika, D.; Karlinasari, L.; Arinana, A.; Batubara, I.; Sitanggang, P.S.; Santoso, D.; Witasari, L.D.; Rachmayanti, Y.; Firmansyah, D.; Sudiana, I.K.; et al. Chemical Components of Fungus Comb from Indo-Malayan Termite *Macrotermes gilvus* Hagen Mound and Its Bioactivity against Wood-Staining Fungi. *Forests* 2021, 12, 1591. <https://doi.org/10.3390/f12111591>
- Naufalin, R., Astuti, S. D., & Wicaksono, R. (2011). Produksi Coating Antimikroba Berbasis Lilin Alami Dan Komposit Pati Dengan Senyawa Antimikroba Ekstrak Limbah Daun Tembakau Untuk Penanganan Pascapanen Buah Dan Sayuran. *Jurnal Litbang Provinsi Jawa Tengah*, 9(2), 160-172
- National Center for Biotechnology Information (2022). PubChem Compound Summary for CID 7041, 2,6-Dimethoxyphenol. Retrieved January 6, 2022 from [https://pubchem.ncbi.nlm.nih.gov/compound/2\\_6-Dimethoxyphenol](https://pubchem.ncbi.nlm.nih.gov/compound/2_6-Dimethoxyphenol).
- Negash, D. A review of aflatoxin: occurrence, prevention, and gaps in both food and feed safety. (2018). *J Nutr Health Food Eng*, 8(2):190 – 197. <https://doi.org/10.15406/jnhfe.2018.08.00268>
- Nerilo, Samuel & Romoli, Jéssica & Nakasugi, Lydiana & Zampieri, Natana & Mossini, Simone & Rocha, Gustavo & da Gloria, Eduardo & Filho, Benício & Junior, Miguel. (2020). Antifungal activity and inhibition of aflatoxins production by *Zingiber officinale* Roscoe essential oil against *Aspergillus flavus* in stored maize grains. *Ciência Rural*. 50. <https://doi.org/10.1590/0103-8478cr20190779>.
- Noviandi, Cuk & Razzazi-Fazeli, Ebrahim & Agus, Ali & Böhm, Josef & Hulan, Hw & Wedhastri, S & Maryudhani, YB & Nuryono, Nuryono & Sardjono, & Leibetseder, J. (2001). Natural occurrence of aflatoxin B1 in some Indonesian food and feed products in Yogyakarta in year 1998–1999. *Mycotoxin Research*. 17. 174-177. <https://doi.org/10.1007/BF03036430>.
- Nweze, E.I., Mukherjee, P.K., Ghannoum, M.A. (2010). Agar-based disk diffusion assay for susceptibility testing of dermatophytes. *Journal of Clinical Microbiology*, Oct 2010, p. 3750-3752. <https://doi.org/10.1128/JCM.01357-10>
- Oakley, K. L., Moore, C. B., & Denning, D. W. (1999). Comparison of in vitro activity of liposomal nystatin against *Aspergillus* species with those of nystatin, amphotericin B (AB) deoxycholate, AB colloidal dispersion, liposomal AB, AB lipid complex,



- and itraconazole. *Antimicrobial agents and chemotherapy*, 43(5):1264–1266.  
<https://doi.org/10.1128/AAC.43.5.1264>
- Passone, M.A., Etcheverry, M.G. (2013). Antifungal impact of volatile fractions of *Peumus boldus* and *Lippia turbinata* on *Aspergillus* section Flavi and residual levels of these oils in irradiated peanut. *International journal of food microbiology*, 168-169C. 17-23. <https://doi.org/10.1016/j.ijfoodmicro.2013.10.009>.
- Parekh, J., Chanda, S. (2008). In vitro antifungal activity of methanol extracts of some Indian medicinal plants against pathogenic yeast and moulds. *African Journal of Biotechnology*, 7 (23), pp. 4349-4353
- Payne, G. A., and Brown, M. P. (1998). Genetics and physiology of aflatoxin biosynthesis. *Annu. Rev. Phytopathol.* 36, 329–362.  
<https://doi.org/10.1146/annurev.phyto.36.1.329>
- Pflieger, W.P., Pócsi, I., Györi, Z., Pusztaheily, T. (2020). The *Aspergilli* and their mycotoxins: metabolic interactions with plants and the soil biota. *Front. Microbiol.* <https://doi.org/10.3389/fmicb.2019.02921>
- Perrone, G., Haidukowski, M., Stea, G., Epifani, F., Bandyopadhyay, R., Leslie, J. F. (2014). Population structure and Aflatoxin production by *Aspergillus* Sect. Flavi from maize in Nigeria and Ghana. *Food Microbiol.* 41, 52–59. <https://doi.org/10.1016/j.fm.2013.12.005>
- Pitt, J., Hocking, A.D. (1999). *Fungi and Food Spoilage*. Maryland: Aspen Publishers, Inc., Gaithersburg,
- Pickova, D., Ostry, V., Toman, J., Malir, F. (2021). Aflatoxins: History, significant milestones, recent data on their toxicity and ways to mitigation. *Toxins*, 13(6): 399. <https://doi.org/10.3390/toxins13060399>
- Pujol, I., Aguilar, C., Fernández-Ballart, J., Guarro, J. (2000). Comparison of the minimum fungicidal concentration of amphotericin B determined in filamentous fungi by macrodilution and microdilution methods. *Med Mycol.* 38(1):23-6. <https://doi.org/10.1080/mmy.38.1.23.26>. PMID: 10746223.
- Qian, Q., Li, S., Wen, H-A. (2011). Fungal diversity of fungus comb in termite nests. *Mycosistema*, 30: 556-565
- Quist, C. F., Bounous, D. I., Kilburn, J. V., Nettles, V. F., Wyatt, R. D. (2000). The effect of dietary aflatoxin on wild turkey poult. *Journal of Wildlife Diseases*, Vol. 36 (3): 436-444
- Rahayu, E.S., (2020). Aspergillus flavus dan Para Saudara Dekatnya.  
<https://cfns.ugm.ac.id/2020/08/15/aspergillus-flavus-dan-para-saudara-dekatnya/>



Rodrigues, M.L. (2018). The multifunctional fungal ergosterol. *mBio*, 9:e01755- 18.

<https://doi.org/10.1128/mBio.01755-18>

Rotimi, O., Rotimi, S., Uchechukwu, Duru, C., Ebebeinwe, O., Obhio Abiodun, A.,

Oluwamayowa Oyeniyi, B. (2017). Acute aflatoxin B1 – Induced hepatotoxicity alters gene expression and disrupts lipid and lipoprotein metabolism in rats. *Toxicol. Rep.* 4, 408–414. <https://doi.org/10.1016/j.toxrep.2017.07.006>

Rouland-Lefèvre, C., Inoue, T., Johjima, T. (2006). Termitomyces/termite interactions.

*Soil Biol.*, 6: 335-350. [https://doi.org/10.1007/3-540-28185-1\\_14](https://doi.org/10.1007/3-540-28185-1_14)

dos Santos, A. G., Marquês, J. T., Carreira, A. C., Castro, I. R., Viana, A. S., Mingeot-

Leclercq, M.-P., Silva, L. C. (2017). The molecular mechanism of Nystatin action is dependent on the membrane biophysical properties and lipid composition. *Physical Chemistry Chemical Physics*, 19(44), 30078–30088.

<https://doi.org/10.1039/c7cp05353c>

Sawhasan P., Worapong J, Flegel TW, Vinijsanun T. (2012). Fungal partnerships stimulate growth of Termitomyces clypeatus stalk mycelium in vitro. *World J Microbiol Biotechnol.* 28(6):2311-8. <https://doi.org/10.1007/s11274-012-1038-x>. PMID: 22806105.

Scheidegger, K. A., Payne, G. A. (2003). Unlocking the secrets behind secondary metabolism: A review of *Aspergillus flavus* from pathogenicity to functional genomics. *J Toxicol* 22, 423–459.

Shakhatreh, M.A., Al-Smadi, M.L., Khabour, O.F., Shuaibu, F.A., Hussein, E.I., Alzoubi, K.H. (2016). Study of the antibacterial and antifungal activities of synthetic benzyl bromides, ketones, and corresponding chalcone derivatives. *Drug Des Devel Ther*, 10:3653-3660. <https://doi.org/10.2147/DDDT.S116312>

Sipos, P., Peles, F., Brasso, D.R., Beri, B., Pusztahelyi, T., Pocsi, I., Gyori, Z. (2021). Physical and chemical methods for reduction in aflatoxin content of feed and food. *Toxins*, 13(3): 204. <https://doi.org/10.3390/toxins13030204>

Stanley, V.C., English, M.P. (1965). Some effects of nystatin on the growth of four *Aspergillus* species. *J Gen Microbiol*, 40:107–118

Subekti, N. (2010). Kelimpahan, sebaran, dan arsitektur sarang serta ukuran populasi rayap tanah *Macrotermes gilvus* Hagen (Blattodea: Termitidaw) di Cagar Alam Yanlappa Jawa Barat [Doctoral dissertation, Universitas Negeri Semarang]. UNNES Library Repository. <http://lib.unnes.ac.id/41965/>



- Subramaniyan, S. & Ganesan, P., Jackson, A., Sandhaanam, S.D., Gabriel P., Michael, Ignacimuthu, S. (2018). Activity of some medicinal plants against phytopathogenic fungi. 5. 124-137. <https://doi.org/10.26438/ijsrbs/v5i5.124137>.
- Sukartana, Paimin. (1995). Natural resistance of 30 wood species against subterranean termite *Macrotermes gilvus* (Hagen); An accelerated field test. *Forest Products Research Journal*, 13(2): 71-76.
- Sukmawati, Ika & Yuniarto, Ari & Rakhmawati, Diah. (2018). Antifungal activity of extract and fraction of Auricularia Auricular on Candida Albicans, Microsporum Gypseum, and aspergillus flavus. *Asian Journal of Pharmaceutical and Clinical Research*, 11. <https://doi.org/10.22159/ajpcr.2018.v11s1.26591>.
- Suzuki, K., Nakano, N., Tanaka, R., Uyeda, M., Shibata, M. 1988. Cell aggregation factor produced by streptomyces sp. Strain No. A-3315. *Agric. Biol. Chem.* 52: 2589–2595.
- Tajehmiri, A., Rahmani, M.R., Moosavi, S.S. (2018). Antifungal effects of six herbal extracts against *Aspergillus* sp. and compared to amphotericin B and nystatin. *International Journal of Advanced and Applied Sciences*, 5(7): 53-57. <https://doi.org/10.21833/ijaas.2018.07.007>
- Takahagi-Nakaira, E., Sugizaki, M.F., Peraçoli, M.T.S. Microdilution procedure for antifungal susceptibility testing of *Paracoccidioides brasiliensis* to amphotericin b and itraconazole. *Journal of Venomous Animals and Toxins including Tropical Diseases* [online]. 2009, v. 15, n. 4 [Accessed 14 November 2021], pp. 718-731. Available from: <<https://doi.org/10.1590/S1678-91992009000400010>>. Epub 27 Nov 2009. ISSN 1678-9199. <https://doi.org/10.1590/S1678-91992009000400010>.
- Uli, A.D.S., Nurtjahja, K., Zuhra, C.F. (2012). Penghambatan pertumbuhan *Aspergillus flavus* dan *Fusarium moniliforme* oleh ekstrak seruni (*Wedelia biflora*) dan kembang bulan (*Tithonia diversifolia*). *Saintia Biologi*, <https://media.neliti.com/media/publications/221080-none.pdf>
- Vidhya, R., and Udayakumar, R. (2017). Antifungal efficacy of leaf, flower, and root of *Aerva lanata* (Linn.) against selected fungal pathogens. *J Immunol Clin Microbiol*, 2(1): 7-13. <https://doi.org/10.5455/jicm.19.20161205>
- Witasari, L.D., Wahyu, K.W., Anugrahani, B.J., Kurniawan, D.C., Haryanto, A., Nandika, D., Karlinasari, L., Arinana, Batubara, I., Santoso, D., Rachmayanti, Y., Firmansyah, D., Sudiana, I.K., Hertanto, D.M. (2021). Antibacterial and antifungal activities of fungus comb extracts isolated from Indo-Malayan termite *Macrotermes gilvus* hagen mound. <https://doi.org/10.21203/rs.3.rs-1159665/v1arifin>



World Health Organization (WHO). (2018). *Aflatoxins*. Department of Food Safety and Zoonoses. REF No: WHO/NHM/FOS/RAM/18.1

Wulansari, D., Praptiwi, Julistiono, H., Nurkanto, A., Agusta, A. (2016). Antifungal activity of (+)-2,2'-epicytoskyrin A and its membrane-disruptive action. *Makara J. Science*, 160-166. <https://doi.org/10.7454/mss.v20i4.6703>

Yamasaki, M., Tamura, N., Nakamura, K., Sasaki, N., Murakami, M., Rajapakshage, W., Kumara, B., Tamura, Y., Lim, S.Y., Ohta, H., Takiguchi, M. (2011). Effects and mechanisms of action of polyene macrolide antibiotic nystatin on Babesia gibsoni in vitro. *J Parasitol*, 97(6):1190-2. <https://doi.org/10.1645/GE-2799.1>. Epub 2011 Jul 14. PMID: 21756054.